

JOURNAL OF PHARMACY AND PHARMACOLOGY

VOLUME XV No. 5

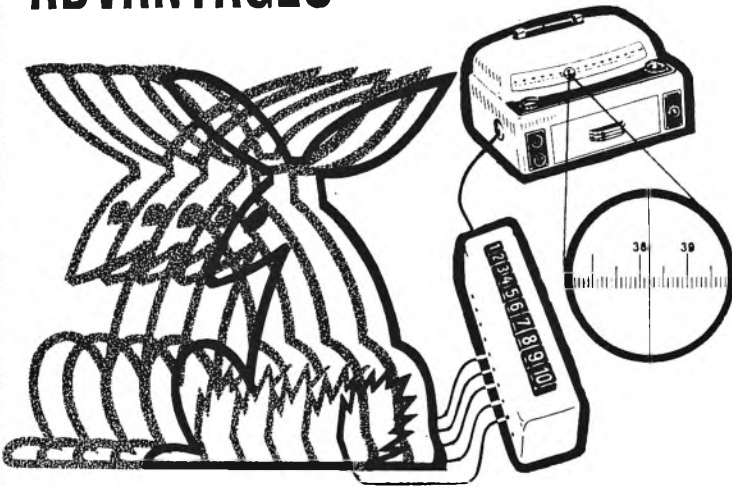


MAY 1963

Published by Direction of the Council of
THE PHARMACEUTICAL SOCIETY OF GREAT BRITAIN

17 BLOOMSBURY SQUARE, LONDON, W.C.1

PYROGEN TESTING with ELECTRIC THERMOMETER TYPE TE3 OFFERS SEVEN IMPORTANT ADVANTAGES



This precision electric thermometer affords distinct and important advantages over the ordinary mercury type. It is used for conducting pyrogen tests by serological institutes, pharmaceutical laboratories and hospitals in more than 30 countries.

1. Guaranteed accuracy to within ± 0.1 centigrade.
2. Easy-to-read spot-illuminated scale.
3. Automatic compensation for variations in room temperature.
4. Up to 30 animals can be tested simultaneously.
5. Temperature indicated within 1 to 2 seconds of depressing push-button.
6. Always ready for immediate use.
7. No tiresome calibration necessary.

SIEREX LTD

*For fully descriptive literature
and prices write to:*

15-18 CLIPSTONE STREET, LONDON, W.1

Tel.: LANgham 2464

JOURNAL OF PHARMACY AND PHARMACOLOGY

Editor: *George Brownlee, D.Sc., Ph.D., F.P.S.*

Assistant Editor: *J. R. Fowler, B.Pharm., F.P.S.*

Annual Subscription £5 0s. 0d. Single Copies 10s.

17 BLOOMSBURY SQUARE, LONDON, W.C.1

Cables: Pharmakon, London, W.C.1. Telephone: HOLborn 8967

Vol. XV No. 5

May, 1963

CONTENTS

Review Article	PAGES
THE RELATION BETWEEN CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY. By J. M. van Rossum, Ph.D.	285-316
Research Papers	
THE CRITICAL MICELLE CONCENTRATIONS OF DOUBLE LONG-CHAIN ELECTROLYTES (AMINE SOAPS) IN AQUEOUS SOLUTION. By A. Packter and M. Donbrow	317-324
THE QUANTITATIVE CONVERSION OF BARBALOIN TO ALOE-EMODIN AND ITS APPLICATION TO THE EVALUATION OF ALOES. By J. W. Fairbairn and S. Simic	325-328
SOME ASPECTS OF THE PHARMACOLOGY OF ORPHENADRINE. By G. Onuaguluchi and J. J. Lewis	329-336
A NOTE ON THE PREPARATION OF RICINOLEIC ACID BY UREA COMPLEXING. By Diptish Chakravarty and Arun Bose	337-338
A NOTE ON THE STABILITY OF THE TRIFLUOROMETHYL GROUP OF BENDROFLUMETHIAZIDE IN RATS. By G. Hasselmann and K. Roholt	339-340
A NOTE ON THE SYNTHESIS OF ESTERS OF <i>N</i> -METHYLPYRROLIDINYL-ALKANOLS. By F. Perks and P. J. Russell	341-343
Letters to the Editor	
MOUSE STRAIN DIFFERENCE IN RESPONSE TO ANTIHISTAMINE DRUGS. By Annie M. Brown	344-345
GAMMA IRRADIATION OF <i>Bacillus subtilis</i> SPORES. By A. M. Cook and T. A. Roberts	345-346
THE ANTIBACTERIAL ACTION OF GLYCINE. By R. V. John and A. D. Russell	346-347
DIFFUSION OF SALTS THROUGH A LIPO-PROTEIN INTERFACE. By L. Saunders	348

EDITORIAL BOARD

H. S. BEAN, B.Pharm., Ph.D., F.P.S., D. C. GARRATT, D.Sc., Ph.D., F.R.I.C., J. C. HANBURY, M.A., B.Pharm., F.P.S., F.R.I.C., F. HARTLEY, B.Sc., Ph.D., F.P.S., F.R.I.C., E. F. HERSANT, B.Pharm., Ph.D., F.P.S., F.R.I.C., J. J. LEWIS, M.Sc., F.P.S., A. D. MACDONALD, M.D., M.A., M.Sc., A. MCCOUBREY, B.Sc., Ph.D., M.P.S., F.R.I.C., D. W. MATHIESON, B.Sc., Ph.D., F.R.I.C., H. G. ROLFE, B.Sc., F.P.S., F.R.I.C., G. F. SOMERS, B.Sc., Ph.D., F.P.S., J. B. STENLAKE, D.Sc., Ph.D., F.P.S., F.R.I.C., G. B. WEST, B.Pharm., D.Sc., Ph.D., F.P.S., R. T. WILLIAMS, D.Sc., Ph.D.

SECRETARY: F. W. ADAMS, B.Sc., F.P.S., F.R.I.C.

DIFCO

microbiological reagents and media

THE ONLY COMPLETE LINE OFFERED IN U.K.

Requirements of the Bacteriologist,
Biochemist, Biologist, Pathologist
and Pharmacologist can usually be
met promptly from our extensive
stocks. We shall always be pleased to
obtain other items specially to order.

*Over 60 years
experience ensure*

**UNIFORMITY
STABILITY
ECONOMY**

Culture Media
Microbiological Assay Media
Tissue Culture Media
Serological Reagents
Antisera
Diagnostic Reagents
Sensitivity Disks
Unidisks
Peptones
Hydrolysates
Amino Acids
Enzymes
Enrichments
Dyes
Indicators
Carbohydrates
Biochemicals

*Please send for
the latest technical
information.*



BAIRD & TATLOCK (LONDON) LIMITED, CHADWELL HEATH, ESSEX, ENGLAND.
Branches in London, Manchester and Glasgow.

REVIEW ARTICLE

THE RELATION BETWEEN CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY*

DISCUSSION OF POSSIBILITIES, PITFALLS AND LIMITATIONS

BY J. M. VAN ROSSUM, Ph.D.

Department of Pharmacology, Catholic University of Nijmegen, the Netherlands

THE number of new compounds being synthesised every year increases in an exponential way. Consequently investigations of the relation between chemical structure and biological activity similarly increases. Studies of this kind provide information about which moieties in the molecule are essential to obtain potent and specific drugs. Furthermore they may give a clue to the mechanism of the action of drugs.

Two major aspects of structure-activity studies raise questions. Firstly, how do alterations in the chemical structure alter the geometry of the molecule and its physical properties? Secondly, what does the fact of biological activity imply? Do different drugs produce the same kind of effect; do they have an identical mechanism of action?

In this review the value of studies on structure-activity, as well as some of the pitfalls and limitations of interpretation will be discussed. Work done on the pharmacology of the nervous system will be used to illustrate this discussion.

ACTION AND ACTIVITY

Investigations of structure activity relationships will be abbreviated to SAR studies. This notation is vague because it is also frequently used for structure action relationship studies.

The action, or effect, of a drug is a qualitative phenomenon while its activity is a quantitative measure denoting how much effect is produced.

When the activity of a series of drugs is compared, as in SAR studies, it is essential that all members of the series cause the identical effect; and not only the same effect, but they also should have an identical mechanism of action. In many examples this is not so.

Different drugs may produce a similar effect by an entirely different mechanism of action. For instance, the blood pressure may be lowered by acetylcholine as a result of vasodilatation of arterioles, but also by histamine when large numbers of capillaries are themselves dilated. It is evident that when the observed effect is achieved by a complicated function, many drugs produce similar effects by acting in different ways while, on the other hand, a single drug may produce a divergency of actions. This is so with drugs acting in the intact animal and especially when they act on the central nervous system. The study of drugs on isolated organs makes things less complicated but it is not always possible to extrapolate *in vitro* findings to intact animals or to man.

* Based on a lecture given for the pharmacological day during the XXII International Congress of Physiological Sciences, Leiden, on September 18, 1962.

Therefore to reduce variables to a minimum the activity must be measured in unambiguous terms. Some knowledge of molecular pharmacology seems necessary for this purpose.

Interaction with Receptors

Most drugs are supposed to interact with receptive sites or receptors in the tissue. The molecular structure of the receptor and its physical and chemical properties largely determine what moieties in the drug are essential for affinity with the receptors. Other factors however also play a role.

Some receptors have strong structural requirements and therefore seem to be rather rigid, whereas others are very much less so. Thus the sites of activity of the volatile anaesthetics are hardly to be called receptors. The potency of these drugs depends largely on their being in the tissue fluid where chlorhydrate formation (Pauling, 1961) may occur, so that the electrical impedance of the tissue increases and conduction is impaired. Ferguson (1930) has calculated the activity, that is, the effective concentration, of volatile anaesthetics at their locus of action and was able to show that they differed little in potency. The geometric structure seems of little importance and there is not a great difference in potency between diastereoisomers and enantiomorphs.

TABLE I
SAR ON SPECIFIC AND NONSPECIFIC RECEPTORS, ACTIVITY ESTIMATED ON THE GUINEA-PIG ILEUM

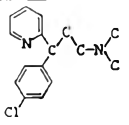
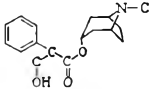
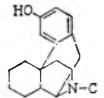
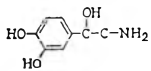
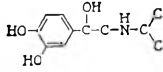

R ₁	R ₂	parasympatholytic (atropine) = 10	histaminolytic (diphenhydramine) = 10	spasmolytic (papaverine) = 1
-C-C-N ^C _C	-H	0.1-1	40-80	1-2
-C-C-N ^C _C	-Cl	0.1-1	10-20	0.5-1
-C-C-O-C-C-N ^C _C	-H	0.1-1	0.1-1	-
	-H	5-10	64-128	-
	-H	80-160	40-80	1-2
-C-C-N ^{C-C} _{C-C}	-H	0.1-10	5-10	0.1-0.5

Other types of drug action involve a requirement for highly structural receptors as, for instance, parasympathetic receptors or histamine receptors. Examples are set out in Table I (Stelt, Harms and Nauta, 1961). From this Table it may be seen that changes in the molecular structure may cause the potency of the antihistamine or antiacetylcholine drug to increase sharply, whereas spasmolytic (papaverine-like) drugs acting on

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

the muscle directly, seem hardly to be influenced. The papaverine-like effects seem to depend on non-specific receptors. For drugs acting on inflexible receptors there may be a great difference in the activity of optical

TABLE II
DIFFERENCE IN AFFINITY OF OPTICAL ISOMERS

drug and (class)	formula	affinity ratio enantiomorphs
chlorpheniramine (antihistaminic)		12 (+)/(−)
hyoscyamine (p-sympatholytic)		100 (−)/(+)
levorphan (analgesic)		150 (−)/(+)
noradrenaline (α-sympathomim.)		3-4 (−)/(+)
isoprenaline (β-sympathomim.)		500 (−)/(+)
nicotine (ganglion stimulant)		equal

isomers. See Table II (Ariëns, 1962; Barlow, 1960; Beckett, 1959; Hanna, 1960; Kroneberg, 1955; Luduena, 1957, 1962; Schild, 1947).

BASIC STEPS IN DRUG ACTION

Since drugs are chemical substances an interaction of drug molecules with specific receptive molecules (receptors) in the biological tissue is an essential part in almost any form of drug action. For receptor occupation a sufficient number of drug molecules should be in the direct vicinity of the receptors. This environment has been called the biophase (Furchgott, 1955). The *first basic step* in drug action is therefore to discover how drug molecules reach the biophase and thus to build up an adequate concentration after administering the drug. It is obvious that this relation depends on the dose, the route of administration, on where the receptors are located in the tissue (on the exterior surface of the cell, or inside it, or in the brain) and on the biotransformation by enzymes.

In intact animals, and especially when drugs are acting in the brain, the relationship between the dose applied and the concentration in the biophase may be very complicated. Furthermore, the concentration in the

biophase is not constant but changes with time as a result of bio-inactivation, excretion and so on. Under standard conditions it is only possible to reach a steady state *in vivo* by infusion techniques.

Obviously the study of the action and activity of drugs on isolated tissue or enzymes is of advantage, since then the concentration in the biophase is more or less directly proportional to the dosage.

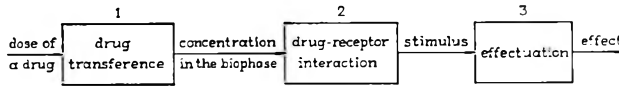


FIG. 1. Block scheme by which the sequence of events leading to the effect by application of a drug may be described. Block 1 represents the relationship between the dose applied and the concentration of the drug in the biophase. The influences of transport, metabolism, excretion etc. are collected in this block and headed under the term drug transference. The reaction of drug molecules with specific receptors under formation of the stimulus is presented in block 2, while the effectuation of the stimulus is given in block 3. From van Rossum (1958).

The *second step* is the drug receptor interaction proper (see Fig. 1). There is good evidence that the reaction between drug molecules and receptors is a bimolecular reaction (Ariëns and van Rossum, 1957b).



where $[R]$ is the concentration of free receptors, $[RA]$ the concentration of occupied receptors and $[A]$ the concentration of drug A. There is no estimate of the total amount of receptors in a tissue so that we can calculate only the fraction of receptors ($[RA]/r$) occupied by the drug:

$$[RA]/r = 1/(1 + K_A/[A]) \quad \dots \quad (2)$$

where r is the total number of receptors and K_A is the dissociation constant of drug A. The affinity of a drug A is defined as the reciprocal of the dissociation constant. The logarithm of the affinity equals the negative logarithm of K_A .

As a consequence of receptor occupation by an agonistic drug a stimulus is generated (Ariëns, van Rossum and Koopman, 1960; van Rossum, 1958; Stephenson, 1956).

$$S_A/S_m = \alpha.[RA]/r \quad \dots \quad (3)$$

where S_A/S_m is the numerical value for the stimulus generated by a certain dose of drug A, while S_m is the maximum possible stimulus to be obtained by the particular tissue. The proportionality constant, α , has been called the "*intrinsic activity*" (Ariëns, 1954; Ariëns, van Rossum and Simonis, 1956a,b).

The intrinsic activity determines whether the drug can generate a stimulus and therefore whether a drug can be "active" or not. The affinity and the intrinsic activity are determined by the molecular properties of both receptor and drug. In structure activity studies these parameters must therefore be determined in the most direct way.

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

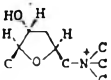

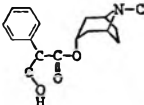
The *third basic step* is the effectuation of the stimulus, that is, the relationship between stimulus and ultimate effect. This relation is not dependent on the drug but determined by the tissue or animal. It is obvious that in isolated organs this relation may be relatively simple and often linear. But, in the intact animal and especially with drugs acting in the brain, the stimulus-effect relationship may be extremely complicated. Furthermore in intact animals there may be a feedback circuit which stabilises the system and counteracts disturbances caused by the drug. This is especially so for endocrine functions and the blood pressure.

In the study of drug action, and therefore also in SAR studies, the effect of a single dose of a drug or the dose which causes a certain effect, is often measured. In certain cases dose-response curves are determined, here, the effects of various doses of the same drug are estimated. The potency of drugs may be calculated from the single dose-technique or from dose-response curves. It should be emphasised that the activity so measured depends on all three basic steps of the scheme in Fig. 1. Therefore it may not be concluded that the potent drugs in a series have the best fit on the receptor unless drug transference to the biophase is identical and the effectuation of the stimulus is a linear function. By using enantiomorphs we often eliminate differences in drug transference but not in effectuation. In such a situation the single dose-technique is not conclusive, since the activity so measured is a function of both intrinsic activity and affinity. These are the limitations of SAR studies and may be the source of large errors when conclusions about the configuration of the receptors are drawn from SAR studies.

Intrinsic Activity

It must be stressed that for SAR studies both the affinity *and* the intrinsic activity should be measured. The intrinsic activity is a measure for the agonistic potencies of a drug. Thus true agonists have a high intrinsic activity whereas competitive antagonists have a low intrinsic activity. This is analogous to substrates and competitive inhibitors in enzymology; the former have a real k_3 value, the latter a low or zero k_3 value (van Rossum and Hurkmans, 1962).

TABLE III
MOIETIES FOR AFFINITY AND INTRINSIC ACTIVITY

			
	muscarine natural	tetramethylammonium	atropine
affinity	high	low	high
intrinsic activity	high	high	low

Different moieties may be responsible for intrinsic activity and affinity. So the conformation is important for affinity in muscarine while the

positively charged ammonium group is important for the intrinsic activity (see Table III). For sympathomimetic drugs the amino-group is important for intrinsic activity on α -receptors but the phenolic OH-groups are essential for intrinsic activity on β -receptors (Ariëns, 1960, 1962; Ariëns and Simonis, 1960).

Stereoisomers often differ in activity (see Table II). This in general is attributed to a difference in affinity, that is, the one substance fits well while its enantiomorphs fit badly with the receptor. However for some drugs the enantiomorphs have similar affinity but different intrinsic activity. This is so for isoprenaline on α -sympathetic receptors (Ariëns, 1962; Luduena, 1962).

The Measurement of Affinity and Intrinsic Activity

We have emphasised that it is important to determine both affinity and intrinsic activity instead of merely a potency or activity. How is such a differentiation made? It cannot be done by the estimation of equiactive doses of drugs; it can be done only by making dose-response curves and, even then, there are pitfalls and restrictions. Dose-response curves can

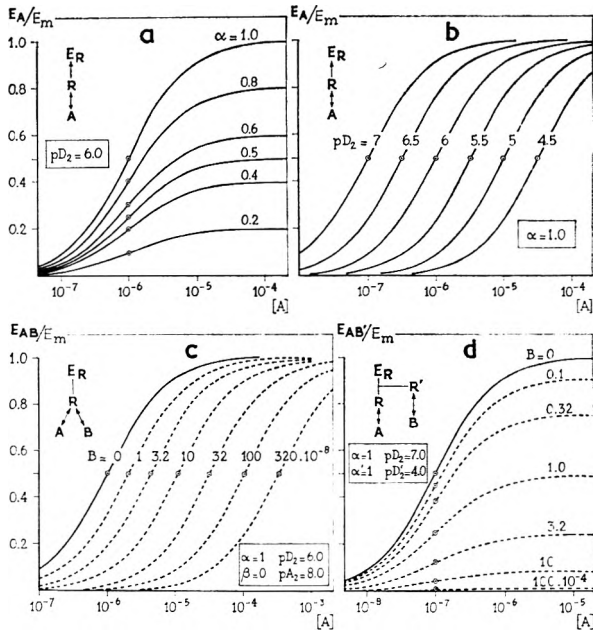


FIG. 2. Theoretical dose-response curves. (a) Dose-response curves calculated for drugs with equal affinity ($pD_2 = 6$) and different intrinsic activities. The maximum height of the curve is directly correlated with the intrinsic activity. (b) Dose-response curves calculated for drugs with equal intrinsic activity ($\alpha = 1$) but different affinities. The position of the curve on the dose axis correlates with the affinity. (c) Dose-response curves for an agonist in the presence of a competitive antagonist. The dose-response curves remain of identical shape and are merely shifted to higher doses along the dose axis. (d) Dose-response curves for an agonist in the presence of different concentrations of a non-competitive antagonist. Dose-response curves are reduced in height whereas the position on the dose axis remains unchanged.

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

only be made for agonists since antagonists are inactive on their own. The maximum height of the dose-response curves is a measure of the intrinsic activity while the position of the curve on the dose axis is a measure of the affinity (see Fig. 2a, b). The logarithm of the affinity ($-\log K_A$) can be calculated as the negative logarithm of the dose which causes 50 per cent of the maximum possible effect. This parameter is denoted the pD_2 value (Ariëns and van Rossum, 1957a).

$$pD_2 = -\log[A]_{50}$$

where $[A]_{50}$ is the concentration of A which causes an effect of 50 per cent (ED50). The intrinsic activity can be calculated only by reference to a standard agonist (S)

$$\alpha = E_{Am}/E_{Sm}$$

where α is the intrinsic activity, E_{Am} is the maximum effect of drug A and E_{Sm} is the maximum effect of the standard drug. It should be noted that the α and pD_2 values so obtained are true values for intrinsic activity and affinity only of an agonist under ideal conditions, that is, when there is a linear relationship of the factors involved in drug transference and

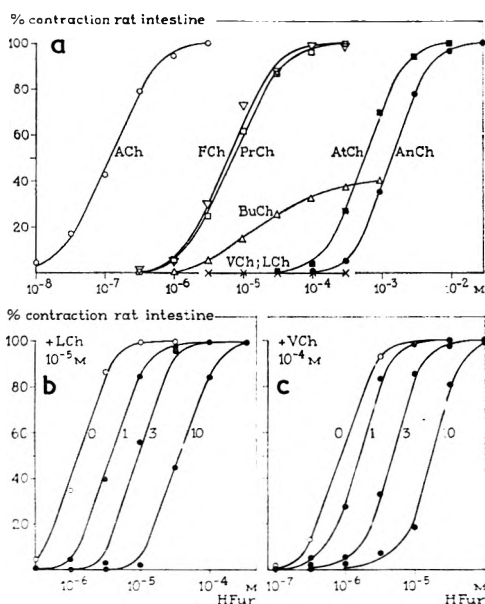


FIG. 3. Dose-response curves for acetylcholine analogues as studied on the isolated rat intestine. (a) Changes in the molecular structure of acetylcholine reflects in changes in affinity and intrinsic activity. Acetylcholine is the most potent drug of this series, whereas acetylthiocholine has a very low affinity but still high intrinsic activity. Butyrylcholine has an intermediate intrinsic activity, valerylcholine and laurylcholine are inactive of their own, Ariëns and Simonis (in the press); Koopman (1963); van Rossum and Hurkmans (1962). (b) Dose-response curves of an agonist in the presence of laurylcholine. The dose-response curves are shifted to higher concentrations along the dose axis indicating a competitive antagonism. (c) Similarly valerylcholine behaves as a competitive antagonist.

similarly with those involved in effectuation (Fig. 1). If, for instance, the stimulus-effect relationship is not linear, the pD_2 may be found too high and the intrinsic activity too low. For details see the literature (Ariëns and others, 1960; Nickerson, 1956; van Rossum, 1958; van Rossum and Ariëns, 1962; Stephenson, 1956).

In Fig. 3, dose-response curves are given for a homologous series of acetylcholine analogues as parasympathetic drugs. In the parent compound, acetylcholine, the methyl group of the acid part is replaced by an alkyl group of increasing length. It may be seen that the affinity decreases gradually but that there is also a gradual decrease in intrinsic activity. The higher homologues are themselves inactive. Formylcholine and acetylthiocholine have a low affinity but high intrinsic activity.

TABLE IV
SAR OF ACETYLCHOLINE DERIVATIVES

formula	drug	α	pD_2	pA_2	affinity ratio
	FCh	1	5.2	—	16
	ACh	1	7.0	—	100
	PrCh	1	5.3	—	2
	BuCh	0.5	5.1	—	1.2
	VCh	0	—	4.7	0.5
	LCh	0	—	5.4	2.5
	AtCh	1	3.3	—	0.02
	AnCh	1	3.0	—	0.01

Ariëns and Simonis, 1963.
Koopman, 1960.
van Rossum and Hurkmans, 1962.

Since antagonists are inactive by themselves they cannot be studied when given alone. Competitive antagonists lack the intrinsic activity but have affinity for the same receptors as the corresponding agonists. They can therefore be investigated in combination with a specific agonist. A parameter comparable to the pD_2 value of an agonist is the pA_2 , as introduced by Schild (1947, 1949) for a competitive antagonist. Under ideal circumstances the pA_2 value is equal to the negative logarithm of the affinity of the competitive antagonist. A competitive antagonist causes

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

a parallel shift of the dose-response curve of a pure agonist (see Fig. 2c). The degree of shifting is a measure of the affinity:

$$pA_2 = -\log[B]_x + \log(x - 1)$$

where $[B]_x$ is the dose of the antagonist which causes a shift of a factor x (Schild, 1957). In Fig. 3b the experimental dose-response curves for a parasympathomimetic and an apparently inactive drug (antagonist) are given. The intrinsic activities and affinities for a series of acetylcholine derivatives are given in Table IV.

Antagonists are not necessarily competitive with the agonist. Some antagonists do not react with the receptors for the agonist but interact with other, although interdependent, receptors. They lack a structural feature that makes the drug fit well on the specific receptors through which the effect of the agonist is brought about. Thus they have their point of action beyond the specific receptors (Ariëns, Simonis and de Groot, 1955; Ariëns, van Rossum and Simonis, 1956b,c) (see block 3 in Fig. 1). These non-competitive antagonists do not cause a shift in the dose-response curves of the agonist but cause a diminution of its maximum height (see Fig. 2d). This property is used to calculate the affinity of the non-competitive drug. A parameter similar to the pD_2 value of an agonist has been denoted the pD'_2 value (Ariëns and van Rossum, 1957).

$$pD'_2 = -\log [B]_{50}$$

where $[B]_{50}$ is the dose of the non-competitive drug that causes a 50 per cent reduction of the dose-response curve of the agonist. Under ideal conditions the pD'_2 value equals the logarithm of the affinity of the non-competitive antagonist. Since the non-competitive drug counteracts the

TABLE V
SAR OF DECAMETHONIUM DERIVATIVES, EXPERIMENTS ON THE ISOLATED RECTUS ABDOMINIS OF THE FROG

R	specific receptors			non-comp receptors	
	α	pD_2	pA_2	α'	pD'_2
-C	0.8	5.9	-	-	-
-C-C	0.3	5.2	-	-	-
-C-C-C	0	-	5.0	-	-
-C-C-C-C	0	-	5.9	-1	5.7
-C-C-C-C-C	0	-	6.2	-1	6.0
-C-C-C-C-C-C	0	-	5.9	-1	5.9
-C-C-C-C-C-C-C	-	-	-	-1	6.2
-C-C-C-C-C-C-C-C	-	-	-	-1	6.4

van Rossum and Ariëns, 1959a.

stimulus which is elicited by the agonists, its intrinsic activity has received a negative sign ($\alpha' = -1$). Intrinsic activities (α and α') and logarithm of affinities (pD_2 , pA_2 and pD'_2 values) for a number of decamethonium derivatives are given in Table V. A differentiation between agonists, competitive and non-competitive antagonists is essential in SAR studies although such drugs may *in vivo* produce a common effect.

AGONISTS AND ANTAGONISTS

Agonists are characterised by the parameters affinity and intrinsic activity but also by the receptors with which they interact (see Table VI). It is generally not possible to isolate receptors, so that in SAR studies the determination of drug parameters on isolated tissues or intact animals must suffice. It should however be ascertained that the various agonists react with the same receptors; this is not proved by the fact that the various agonists cause identical effects. One way to solve this problem is the use of a specific competitive antagonist in combination with the agonist. A specific competitive antagonist should cause the same degree of antagonism in a certain dose, irrespective of the agonist used. In isolated organs such a procedure can be followed but *in vivo* it may not be possible to do so.

TABLE VI
PARAMETERS FOR AGONISTS AND ANTAGONISTS

type of drug	receptor type	affinity	intr. act.
agonist	specific	pD_2	$\alpha = 1$
comp. antagonist	specific	pA_2	$\alpha = 0$
non-comp. antagonist	other (interdependent)	pD'_2	$\alpha' = -1$

In studying different histaminomimetic drugs, receptor identity could be proved by combining the agonists with a selective antihistamine drug (Schild, 1957). Irreversibly acting drugs may also be used to evaluate the receptor identity of agonists (van Rossum, 1958; van Rossum and Ariëns, 1962).

Different agonists may react with identical receptors but they also may reach an identical effect by reacting with different receptors. This means that different agonists may not have an identical mechanism of action. Drugs that cause an identical effect by reacting with different receptors are of a separate class and may be called functional synergists (Ariëns and others, 1956c; van Rossum, 1951; van Rossum and Ariëns, 1959b). Parasympathomimetic drugs and histaminomimetic drugs are functional synergists on the isolated guinea-pig intestine. A differentiation between functional synergists and competitive agonists can be obtained by using selective irreversibly-acting drugs. This implies that one kind of receptor can be blocked permanently whereas the other kind is not affected.

The picture of agonists may further be complicated by drugs acting indirectly by causing the release of an endogenous agonist. The dose-response curves of indirectly-acting drugs are not shifted to the right to the same degree by a selective competitive antagonist as those of directly acting drugs.

When inactive compounds are found in a series of agonists, this may mean either that the intrinsic activity is zero, or that the affinity is nil. If *in vitro* the affinity is very low, the compound does not show any effect. *In vivo*, however, this may not be so since it may have affinity with other receptors. If the intrinsic activity is zero, a drug is only apparently

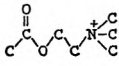
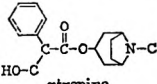
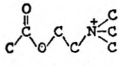
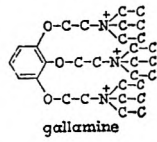
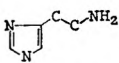
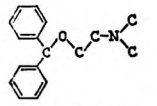
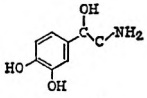
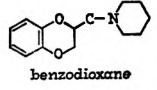
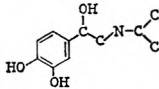
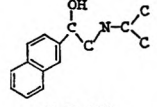
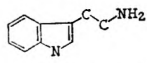
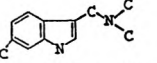
CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

ineffective, since, if the drug still has affinity, it will interfere with receptor occupation by an agonist and thus show competition. In the SAR of the drugs shown in Fig. 3 and Table IV there is a change from agonist to competitive antagonist. This is a common situation when alterations are made in the molecular structure of an agonist.

An inactive drug which has no affinity to the specific receptors may, however, have affinity to non-specific (interdependent) receptors and so act as a non-competitive antagonist when combined with an agonist.

It must again be emphasised that "inactive" drugs are not necessarily inactive. They may be really inactive but, on the other hand, they may be competitive antagonists, non-competitive antagonists or functional agonists in addition. If therefore in a series of drugs, inactive members are encountered they should be studied as antagonists in combination with an appropriate agonist.

TABLE VII
SIZE OF AGONIST AND ANTAGONIST MOLECULES

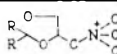


agonist	antagonist	receptor
 acetylcholine	 atropine	parasympathetic
 acetylcholine	 gallamine	myoneural
 histamine	 diphenhydramine	histaminic
 noradrenaline	 benzodioxane	α-sympathetic
 isoprenaline	 Nethalide	β-sympathetic
 5-hydroxytryptamine	 6-methylgramine	5-HT

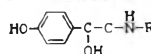


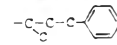
Agonists are usually molecules of smaller size than the corresponding competitive antagonists (see Table VII). While the affinity of agonists is often not very high, competitive antagonists, which do not need moieties

J. M. VAN ROSSUM

for intrinsic activity, can be built for optimal affinity. There being competition, competitive antagonists interact with the same receptors as the corresponding agonist. Actually they do not react with identical receptors. The receptor area used by the antagonist is often larger. It is therefore dangerous to make pictures of the specific receptors for SAR studies of antagonists. A receptor supposed to be complementary to natural muscarine is quite different from the complementary part of hyoscyamine.

TABLE VIII
GRADUAL CHANGE FROM AGONIST TO ANTAGONIST VIA INTERMEDIATES WITH LOW AFFINITY

 parasympathetic receptors (rat intestine)			
R	α	pD ₂	pA ₂
H-	1	5.3	—
C-	1	4.3	—
C-C-	0.5	4.5	—
 -	0	—	4.4
C-C-C-	0	—	6.2
C-C-C-C-	0	—	6.9
 -	0	—	7.1

 α -sympathetic receptors (rat v. deferens)			
R	α	pD ₂	pA ₂
H	1	4.6	—
C	1	4.1	—
C-C-C	0.5	3.0	—
C-C-C-C	—	—	< 2.0*
 -	0.2	—	2.5
 -	0	—	5.6
 -	0	—	5.5

* shows non-competitive antagonistic properties

Interpretation of in vivo experiments

Thus it is important to know whether drugs act as agonists or as antagonists. In isolated organs this is usually not a problem, but in intact animals, and especially with centrally-acting drugs, this may be extremely difficult to determine. For centrally-acting drugs we often do not know whether a stimulant drug really is an agonist or whether it is an antagonist. Strychnine for instance is considered a stimulant drug. It causes inhibition at inhibitory synapses (Eccles, 1961; Longo and Chiavarelli, 1962) but we do not know whether this inhibition is an agonistic action or whether there is competition with a transmitter so that its action would be antagonistic. A number of compounds of less

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

complicated structure have been synthesised (Chiavarelli and Setting, 1958) and screened for true strychnine-like action (Longo and Chiavarelli, 1962). These compounds have not been proved to be truly strychnine-like, while the inactive compounds do not yet seem to have been investigated for antagonistic properties. Studies along these lines are, however, promising.

A similar situation exists with morphine. Is the analgesic effect of morphine an agonistic or an antagonistic action? One might argue that presumably it is agonistic because *N*-allylnormorphine is a morphine antagonist. These drugs are mutual antagonists. We have seen that in most cases antagonists are obtained when larger substituents are introduced. It would then seem likely that morphine is an agonist.

It must be evident that, when larger substituents are introduced in drugs which are already antagonists, the derivatives remain antagonists. It is therefore not logical to anticipate that *N*-allylnoratropine would be an antagonist of atropine. Needless to say, a certain group in one type of drug may produce a totally different effect in another class of drugs.

The fact that competitive drugs are generally of a larger size than the agonists may explain why different series of antagonists, although being competitive with the same agonist, have entirely different requirements for optimal affinity. The different kinds may have affinity with different regions adjacent to the receptor for the agonist. This also may explain why in agonistic molecules small alterations, decreasing the intrinsic activity, also cause a decrease in affinity, while the introduction of aromatic (flat) ring systems increases the affinity again (see Table VIII). The weakest drugs of such a series may show non-competitive antagonistic properties, that is, they may react with other receptors and so interfere with the action of the agonist. The more potent agonists or antagonists may also have non-competitive affinity as well, but in low doses they are specific agonists or antagonists and at high doses they show additional effects. In fact most drugs have a multiple mode of action but they are selective when the ratio between affinity for the desired and undesired receptors is high. Drugs which have a very high affinity for certain kinds of receptors are, in general, specific. This is so since they fulfil the requirements for the particular kind of receptors and thus have little chance of being optimal for another kind, unless the two are strongly interrelated (see also Table I).

TISSUE AND SPECIES DIFFERENCES FOR DRUG PARAMETERS

The drug parameters, affinity and intrinsic activity, are determined by the molecular configuration and physical properties of the drug as well as those of the receptor. Specific receptors, as, for instance, parasympathetic receptors, may differ for different tissues of the same animal and similar tissues of different species. This may reflect in tissue and species differences with respect to affinity and intrinsic activity.

Van Rossum and Ariëns (1959c) have reported on a homologous series of dioxolanes. These parasympathetic drugs were studied on the isolated rat intestine, the isolated frog heart and on the blood pressure of the

anaesthetised cat. With the introduction of heavier substituents there was a decrease in the intrinsic activity so that the lower members which were agonists, gradually changed into competitive antagonists. On the frog heart the transition from agonist to antagonist took place earlier than on the intestine while on the blood pressure of the cat the drugs remained agonists still longer. Thus certain members of the series were pure agonists on one tissue but pure antagonists on another.

Similar results have been obtained for neuromuscular blocking agents on the frog rectus abdominis on the one hand and the chicken muscle on the other hand. Some of these drugs are depolarisers on the chicken but pure competitors on the frog rectus (van Rossum and Ariëns, 1959a).

The transition compounds are of special interest in SAR studies. When the action of a homologous series is studied the variable effect of the transition compounds can be understood. However, when such an intermediate drug is investigated alone, it is difficult to classify because of the variable response in different tissues of the same animal and similar tissues of different species. A nice example is the action of pilocarpine. Pilocarpine is considered to be parasympathomimetic but owing to its relatively large molecular size the intrinsic activity can hardly be as great as that of acetylcholine. In fact, on the intestine, pilocarpine was found to have an intrinsic activity of about 80 per cent of that of acetylcholine (van Rossum 1960b). Since on the heart the intrinsic activity is found to be lower, pilocarpine behaves largely like atropine. When studied on the pupil of the eye extremely variable results may be obtained. Depending on the amount of endogenous acetylcholine released from nerve endings, which causes the pupil to constrict, to a certain degree pilocarpine may either cause a miosis or a mydriasis. On a pupil largely constricted it acts like atropine, while on some animals pilocarpine may even be inactive.

MOLECULAR CONFORMATION AND PHARMACOLOGICAL ACTIVITY

Flexible drug molecules may acquire many possible conformations. Depending on the flexibility of the receptor, sometimes a few, sometimes more, drug conformations may give optimal affinity. Rigid drug molecules which have the correct configuration may have a perfect fit. However in such an example small alterations may reflect great differences in affinity.

There is good evidence that, for instance, parasympathetic and sympathetic receptors are rigidly built, whereas the cholinergic receptors in autonomic ganglia seem to be rather flexible (see below). For rigid receptors there will be a strict SAR and there will then exist a great difference in activity of optimal isomers. If the one isomer fits in an optimal way its enantiomorph does not fit or hardly fits. Table II gives a few examples.

If the enantiomorphs of a drug are both antagonists, a difference in total activity is due to only a difference in affinity. But, for agonists, the enantiomorphs may differ in affinity as well as in intrinsic activity. Since the geometry of the molecule is important for receptor occupation an activity difference for enantiomorphs is for a great part attributed to

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

affinity, while a difference in intrinsic activity may be an additional factor. It is unlikely that the enantiomorphs have identical affinity but differ only in intrinsic activity.

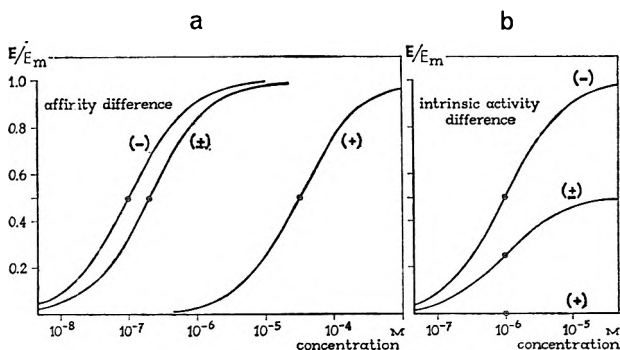


FIG. 4. Theoretical dose-response curves for enantiomorphs and the racemate. (a) The (-)-isomer is considered to have an affinity 300 times higher than the (+)-isomer, while both are considered to have identical intrinsic activity. In such a case the racemate seems to have an affinity of half that of the (-)-isomer. In the racemate the (+)-isomer does not contribute to the effect because of the high potency of the (-)-isomer. (b) The (-)-isomer is considered to have a high intrinsic activity ($\alpha = 1$), whereas the (+)-isomer is considered to have a low intrinsic activity ($\alpha = 0$). Both isomers have equal affinity. The racemate then seems to have an intrinsic activity in between, because equal amount of receptors are occupied by an agonist (-) and by competitive antagonist (+). This remains so over the dose-response curve.

In Fig. 4 dose-response curves are given for (+)-, (-)- and the racemate of theoretical compounds. In the left figure it is assumed that the affinity of the (-)-compound is 300 times higher than that of the (+)-isomer while the intrinsic activities are identical. The racemate is then practically half as potent as the (-)-isomer. In the right figure it is assumed that both isomers have identical affinity but that the (-)-compound has a high intrinsic activity and therefore is a pure agonist, while the (+)-isomer has a low intrinsic activity and therefore is inactive by nature. Actually it acts as competitive antagonist of its enantiomorph. The racemate then seems to have an intrinsic activity in between, which is due to a competition between the enantiomorphs. Since both isomers have equal affinity, at every dose level of the racemate they occupy equal amounts of receptors. That implies that the (-)-isomer can occupy only maximally 50 per cent of all receptors and therefore produce only 50 per cent effect. Obviously a drug with an intrinsic activity equal to 0.5 can also maximally produce 50 per cent effect but in this example all receptors are occupied while the contribution to the effect per occupied receptor is only half of that of a pure agonist.

There are many examples of agonists which differ only in affinity. For instance (+)- and (-)-muscarine and (+) and (-)-methacholine on parasympathetic receptors (Ellenbroek and van Rossum, 1960; Waser, 1958, 1961) and (+)- and (-)-adrenaline on α -sympathetic receptors (Ariëns, 1960, 1962; Hanna, 1960) (see Fig. 5). There are only a few

examples of enantiomorphs with equal affinity but with a difference in intrinsic activity. This is so for (–)- and (+)-isoprenaline (Ariëns, 1962; Luduena, 1962). The racemic mixture apparently has an intermediate intrinsic activity (Ariëns, 1962) (see Fig. 5c).

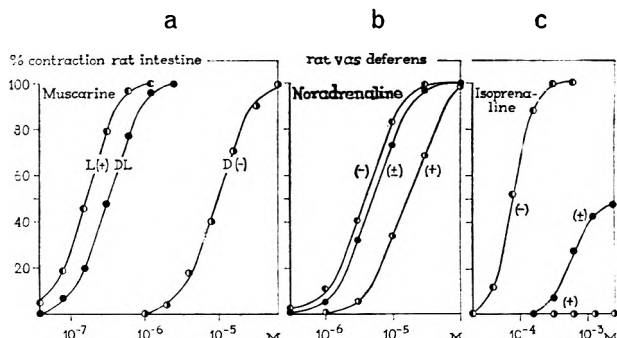


FIG. 5. Dose-response curves of enantiomorphs and their racemate. (a) Enantiomorphs of muscarine as parasympathomimetics on the isolated rat intestine. The L(+)-isomer is twice as potent as the racemate and about 300 times more potent than the D(-)-isomer. The difference in potency is attributed to a difference in affinity. The isomers have equal intrinsic activity (Ellenbroek and van Rossum, 1960). (b) The enantiomorphs of noradrenaline as α -sympathomimetics of the vas deferens of the rat. The (–)-isomer is slightly more potent than the racemate and about 4 times more potent than the (+)-isomer. The different potencies are attributed to differences in affinity. The intrinsic activities are identical. (c) The enantiomorphs of isoprenaline as α -sympathomimetic drugs. The (–)-isomer is an agonist whereas the (+)-isomer is inactive of its own. The racemate shows an intermediate intrinsic activity. The differences in potency are largely attributed to differences in intrinsic activity (Ariëns, 1962).

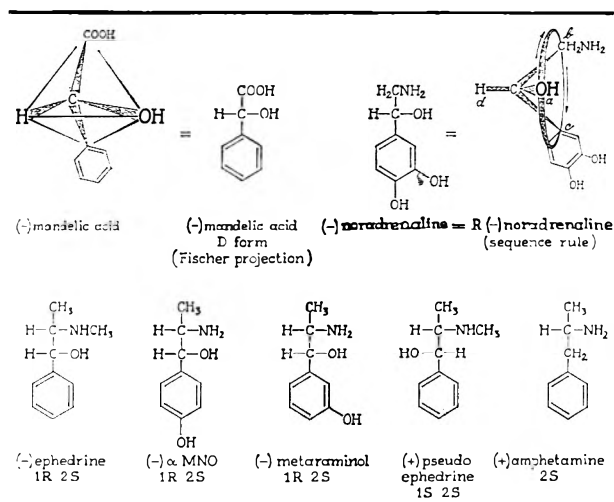
SAR studies of series of enantiomorphs may provide valuable information on the configuration of the receptor since the drugs with optimal fit are more or less complementary with the receptor. Then obviously the absolute configuration of the various active isomers should be known. The optical rotation does not give information about the absolute configuration.

An unambiguous notation of absolute configuration should be used in publications otherwise an enormous amount of confusion will ensue. The Fisher notation D and L seems accurate only for α -amino-acids and α -hydroxycarbonic acids which contain only one asymmetric centre. The sequence rule as introduced by Cahn and Ingold (1951) and Cahn, Ingold and Prelog (1956), meets a need. Knowledge of the sequence rule is a necessity for studying the relationship between geometrical structure and biological activity. In Table IX the absolute configuration of a number of sympathetic drugs is given according to the sequence rule, while furthermore the relationship between the Fisher projection and sequence rule is briefly outlined. The notation R (rectus) and S (sinister) is found as follows. The four substituents around the asymmetric centre are arranged according to their weight (sequence a, b, c, d) for which rules have been given (Cahn and Ingold, 1951; Cahn, Ingold and Prelog, 1956). The group of lowest weight (d) is placed in the

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

axis of a steering wheel. The three other groups complete the steering wheel. The wheel is then rotated in the direction from $a \rightarrow b \rightarrow c$. If such a rotation is to the right the configuration is *R*, and when it is to the left

TABLE IX
NOTATION OF ABSOLUTE CONFIGURATION



Pratesi and others, 1958, 1959.
Dijk and Moed, 1961.
Dirkx, 1962.

left the configuration is denoted as *S*. For details see Cahn and others (1951, 1956). The relationship between absolute configuration and affinity and intrinsic activity of muscarinic drugs is given in Table X.

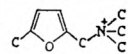
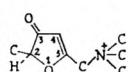
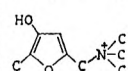
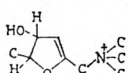
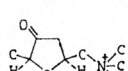
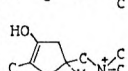
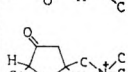
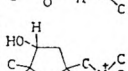
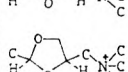
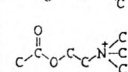
It has been proved by Pratesi, La Manna, Campiglio and Ghislandi (1958, 1959) that natural noradrenaline (laevorotatory) and natural adrenaline have the *R*-configuration. For closely related compounds such as $(-)$ - β -hydroxytyramine and $(-)$ -noradrenaline an optical rotation of the same sign may indicate identical configuration although it is not a proof. For metaraminol (*m*-hydroxynorephedrine) which is also laevorotatory not even a suggestion may be made that the corresponding OH group would have the same configuration as natural adrenaline because two asymmetric carbon atoms are involved (see Table IX).

Rotation dispersion determinations may help in ascertaining the absolute configurations of similar drugs (Dirkx, 1962; Lyle, 1960). It could be proved with rotation dispersion that $(-)$ α -methylnoroxedrine, α -MNO (Table IX) has identical configuration with $(-)$ -ephedrine (Dirkx, 1962). This conclusion had been reached by van Dijk and Moed (1959, 1961) by using chemical methods. By using rotation dispersion techniques Dirkx (personal communication) has recently established the absolute configuration of $(-)$ -metaraminol and $(-)$ -cobefrine (3,4-dihydroxynorephedrine) as 1*R*: 2*S*.

J. M. VAN ROSSUM

When alterations are made in drug molecules changes in both steric factors and in other physical properties are involved. Since the chemical and physical properties of enantiomorphs are largely identical, the only differences are steric features. In general, it may be assumed that the factors of drug distribution are identical for enantiomorphs so that in most instances both isomers acquire the same concentration in the biophase. This implies that if we cannot avoid the biophase relations, as in intact animals, it is useful to work with stereoisomers. Obviously this is not so if active transport or asymmetric metabolism is concerned.

TABLE X
CONFIGURATION AND ACTIVITY OF MUSCARINICS

drug	formula	configuration	form studied	α	pD ₂	relative potency
methyl-furmethide		planar		1	7.2	160
dehydro-muscarone (cis)		2S	(±)	1	7.6	400
tautomer		planar		1		
dehydro-muscarine (trans)		2S : 3R	(±)	1		100
muscarone		2S : 5S	(-)	1	7.8	700
tautomer		5S		1		
allo-muscarone		2R : 5S	(±)	1		170
muscarine		2S : 3R : 5S	(+)	1	7.1	120
methyl-dilvasene		2S : 5S	racemates	1	7.3	200
ACh		-	-	1	7.0	100

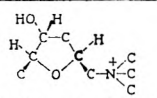
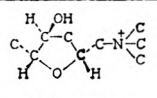
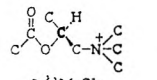
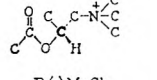
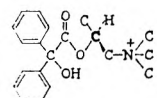
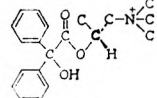
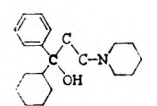
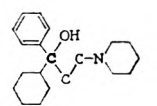
Gyermek and Unna, 1960.
van Rossum and Ariëns, 1959b.
Waser, 1958; 1961.

Laarhoven, Nivard and Havinga (1961) have recently studied stilboestrol derivatives in which effects other than steric effects were minimised. They found that steric hindrance to coplanarity of dimethoxystilbenes is one essential factor for oestrogenic activity. Angles of twist larger than 60° sharply increase activity.

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

It has been pointed out by Pfeiffer (1956) that optical isomers with high activity show a great difference in activity of the isomers. This rule seems to hold for all possible kinds of drugs. Evidently this rule can hold only if the asymmetric centre in the molecule is in a key position with respect to affinity or intrinsic activity (see Table XI). From this Table it may be seen that for the optical isomers of the potent parasympathomimetics muscarine and methacholine there is a great difference in affinity. Both the active muscarine and methacholine have the same configuration (Beckett, 1962; Ellenbroek and van Rossum, 1960). The corresponding asymmetric carbon atom is in a key position.

TABLE XI
KEY POSITION AND ASYMMETRIC CENTRE

enantiomorphs		affinity ratio
 L-(+)-muscarine (natural)	 D-(-)-muscarine	$L/D=300$
 L-(+)-MeCh	 D-(-)-MeCh	$L/D=300$
 L-(+)-BIP	 D-(-)-BIP	$L/D=1$
 (-)-BIP	 (+)-BIP	$(-)/(+)=150$

Ellenbroek and van Rossum, 1960.
Ellenbroek, 1963.
Long and others, 1956.

Now if, instead of the acetyl ester from *S*- and *R*- β -methylcholine, an ester is made with benzylic acid, potent atropine-like drugs are obtained (Ellenbroek, 1963). The high affinity of these esters is due to the phenyl rings which may form van der Waals' binding with an appropriate part in, or near, the receptor. Now there is practically no difference between the enantiomorphs. The asymmetric carbon atom is not in a key position (see Table XI). If however the asymmetric centre is located on the negative site of the molecule again a great difference in affinity of the enantiomorphs is found (Long, Luduena, Tullar and Lands, 1956). The rule of Pfeiffer is valid when the receptors are inflexible molecules and the asymmetric centre is in a key position. Not only for enantiomorphs does it hold that the difference between them is great when the potency is high. It also holds in a more general way for the specificity

of the drug. A drug with an optimal fit on one kind of receptor will, in general, fit badly on other receptors. This is because requirements for the one differ from those of another. Drugs with a low affinity and having a poor fit on specific receptors, but having similar affinity for various sorts of receptors, are therefore not selective. It will be found that small alterations in potent drugs may reflect great variations in drug parameters whereas alterations in the molecule of weak drugs hardly elicit a change.

The considerations which have been discussed will now be applied to SAR studies of some selected classes of drugs acting on the peripheral and central nervous system.

PARASYMPATHOMIMETIC AND ANTI-ACETYLCHOLINE DRUGS

Parasympathomimetic drugs are related to acetylcholine in their molecular structure. They have affinity to specific receptors in cholinergic synapses and in addition a real intrinsic activity. They therefore are often called cholinomimetics. Those which have only affinity are sometimes called cholinolytics.

In the peripheral nervous system, three types of cholinergic receptors can be distinguished. They are the postganglionic parasympathetic, the ganglionic and myoneural receptors.

TABLE XII
SAR OF POTENT PARASYMPATHOMIMETICS

formula	R	drug	α	pD ₂	affinity ratio
	H- C-	FCh ACh	1 1	5.2 7.0	1.6 100
	H- C-	HF MeF	1 1	5.3 7.3	1 100
	H- C-	(±) trans HMu (±) Mu	1 1	4.8 6.8	1 100
	H- C-	(±) H-muscaron (±)-muscaron	1 1	5 7.6	2.5 100
	H- C-	HFur MeFur	1 1	5.9 7.2	6 100

Koopman, 1960.
van Rossum and Ariens, 1959a.
van Rossum and Hurkmans, 1962.

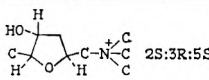
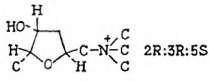
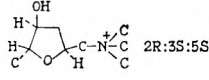
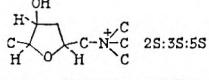
The parasympathetic receptors seem inflexible and consequently there is a strict structure-activity relationship. The agonists on parasympathetic receptors are very similar in molecular configuration (see Table XII). There is a great difference in potency of enantiomorphs.

It is evident that conclusions about the structure of the parasympathetic receptors may not be drawn from SAR studies in which only the overall

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

activities of the structural analogues are compared. A frequently used test for parasympathomimetics (muscarinics) is reduction of the amplitude of the frog heart set up according to Straub. Obviously the heart can be stopped beating in a number of ways. Using as a basis the dose which causes a 25 per cent reduction of the heart amplitude it may be concluded that the four stereoisomers of muscarine differ greatly in parasympathomimetic activity (see Table XIII) (Waser, 1961). The epi-isomer is the weakest muscarinic drug. However, when both affinity and intrinsic activity are determined, it becomes evident that the epi-isomer is largely an atropine-like substance (van Rossum, 1960a). So the activity determined in the "usual" way may lead to incorrect conclusions. From this series of compounds it may be concluded that minor changes in the molecular configuration may cause a change from agonist to antagonist.

TABLE XIII
ACTIVITY OF DIASTEREISOMERS OF (\pm)-MUSCARINE AS ESTIMATED ON THE ISOLATED FROG HEART

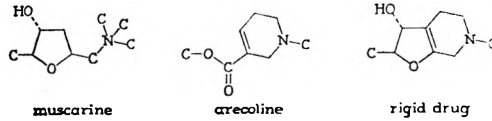
formula and configuration	drug	relative activity	α	pD ₂	pA ₂
 2S:3R:5S	(\pm)-Mu	100	1	6.4	—
 2R:3R:5S	(\pm)-epi- allo-Mu	0.27	1	4.5	—
 2R:3S:5S	(\pm)-allo-Mu	0.21	0.4	—	3.7
 2S:3S:5S	(\pm)-epi-Mu	0.015	0.1	—	3.8

van Rossum, 1960a.
Waser, 1958.

When determining both affinity and intrinsic activity, a distinction can be obtained about which moieties are responsible for affinity, that is, for the binding capacity with the receptor, and which moieties are responsible for the intrinsic activity, that is, for the capacity to act as a mimetic. The quaternary nitrogen atom is essential for the mimetic action of muscarinic drugs while the conformation or configuration is important for affinity. Introduction of heavier substituents on the nitrogen atom causes a gradual change from parasympathomimetic to anti-acetylcholine properties. As a rule, the affinity decreases simultaneously with the intrinsic activity, unless planar ring systems are introduced which increase affinity by combining with adjacent regions near the receptor proper. Compare the structure of atropine and other potent antiparasympathomimetics with the mimetics or the antagonist obtained by gradually introducing heavier substituents in a mimetic drug.

Muscarinic Sites

The potent muscarinic drugs have a rather rigid structure (see Tables X, XII). The quaternary nitrogen atoms of this molecule, however, may still acquire many different conformations. In arecoline, which is also a potent muscarinic drug, the nitrogen atom is in a fixed position in the ring but in this molecule the negative site or the ester configuration may



acquire different conformations. It seems tentative to speculate that in muscarine the nitrogen atom may be in a configuration similar to that in arecoline while in arecoline the ester grouping may in a similar way be held as the negative side of muscarine. Combining these two configurations, an extremely rigid molecule may be built, which should then be a potent muscarinic drug. This awaits further confirmation. The conclusion must be that unless the conformation of a drug when it is on the receptor is known, it is still difficult to draw conclusions about the receptor being complementary to the drug.

TABLE XIV
SAR OF POTENT PARASYMPATHOMIMETICS

agonist quaternary	rat intestine		agonist tertiary	rat intestine	
	α	pD ₂		α	pD ₂
	1	7.0	nor-ACh	1.0	4.5
	1	6.9	nor-MeCh	0.9	3.9
	1	7.0	nor-CarbCh		
	1	7.3	nor-MeFMes	1.0	4.1
	1	7.1	nor-muscarine	0.9	3.8
	1	7.2	nor-MeFurMes	0.8	4.3
	0.9	4.8	arecoline	1.0	6.5

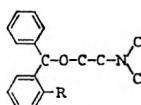
The nitrogen atom in arecoline is tertiary. If this nitrogen atom is made quaternary the intrinsic activity as well as the affinity decreases. This in contrast to acetylcholine and muscarine (see Table XIV). When

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

these compounds are made tertiary there is a tremendous decrease in affinity and intrinsic activity. That the nitrogen atom in arecoline is in the ring is not the reason, since very closely related analogues of arecoline have been made (Jaramillo, 1962). For these analogues the quaternary compound was always more potent than the tertiary. The abnormal behaviour of arecoline has not yet been explained. Atropine-like drugs of other antimuscarinic drugs do not need moieties for intrinsic activity. It is therefore irrelevant whether the nitrogen is tertiary or quaternary (this is only important as far as biophase relations are concerned). Optimal affinity seems to be achieved in different ways.

TABLE XV

2-ALKYLDIPHENHYDRAMINES, ACTIVITY ESTIMATED ON THE ISOLATED GUINEA-PIG ILEUM



R	parasympatholytic activity (affinity)	histaminolytic activity (affinity)	ratio P/h
-H	1	1	1
-C	2.1	0.2	10
-C-C	4.2	0.1	42
-C-C-C	4.9	0.1	49
	6.5	0.1	65
-C-C-C-C	5.5	0.07	79
	16.0	0.1	160
	33.0	0.05	660
laevo	50.0	0.06	830
dextro	0.3	0.06	5

Harms, 1956.




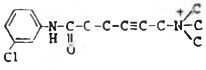
Since anti-muscarinic drugs have no intrinsic activity the potency is a reasonable measure for affinity. In Table XV the SAR is given of a number of diphenhydramine derivatives (Harms, 1956). It must be concluded that substituents which prevent free rotation of the planar rings favour atropine-like properties and that there is a great difference in potency of optical isomers.

Ganglia

The cholinergic receptors in the ganglionic synapse seem to be more flexible. There is no such structure activity relationship (see Table XVI). In experiments where ganglionic stimulants were combined with a selective competitive antagonist, only 1,1-dimethyl-4-phenylpiperazinium iodide (DMPP) and nicotine reacted with common receptors; the nicotine homologue 3-pyridylmethyltrimethylammonium bromide, PMTM, reacted

at least with a partially different receptor (van Rossum, 1962). There was also no difference in activity of stereoisomers. Barlow (1960) has shown that *S*- and *R*-nicotine have identical activity. The cholinergic receptors in the autonomic ganglia differ to a great extent from those in the parasympathetic synapse. The potent and inflexible parasympathomimetics such as muscarine and dioxolanes are only weak ganglionic stimulants, whereas the flexible muscarinics acetylcholine and carbachol have strong ganglionic stimulating properties.

TABLE XVI
SAR OF GANGLIONIC STIMULANTS

formula	drug	guinea-pig intestine	
		α	pD ₂
	nicotine	0.7	5.3
	PMTM	1.0	5.9
	DMPP	0.6	5.6
	MeN-A 343	0.9	4.8

van Rossum, 1962.

Skeletal-myoneural Junction

The cholinergic receptors in the myoneural junction seem to be similar to those in the ganglia but different from the parasympathetic receptors. In the myoneural junction the structure activity relationship among mimetics is not strict. But the nitrogen atom is important for the intrinsic activity in all cholinergic drugs. Introduction of heavier groups on the nitrogen atom of decamethonium, for instance, causes a gradual change from mimetic to lytic (see Table V). This implies a gradual change from a depolarising neuromuscular blocking agent into a non-depolarising competitive neuromuscular blocking agent. When curare-like compounds are studied with the head-drop technique, such a structure activity relationship would not become evident. Using the head-drop technique in SAR studies may suggest that there is no relationship at all between structure and action. However, as previously pointed out, we can speak of a structure-activity relationship only when the drugs under study exert the same mechanism of action. When in decamethonium one methyl group is replaced by an alkyl chain of increasing length, there is a gradual change from mimetic to lytic, but, by further increasing the chain length, the non-depolarising drugs change their mechanism of action; for instance, the heptyl derivative is a pure non-competitive antagonist of acetylcholine (see Table V) (van Rossum and Ariëns, 1959). This compound shows affinity for different receptors and in such a way blocks neuromuscular

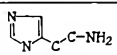
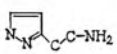
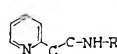
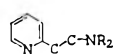
CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

transmission. This clear structure-activity relationship did suggest immediately that a new clinical neuromuscular blocking agent, dioxahexadekanium bromide (G25178), might be of the non-competitive type. Indeed difficulties were encountered in classifying this compound, since apparently it did not belong to the depolarisers nor to the non-depolarising competitors. Actually it appeared to be a pure non-competitive antagonist of acetylcholine and therefore is a nice example of a new type of neuromuscular blocking agent (van Rossum, Ariëns and Linssen, 1958).

HISTAMINOMIMETICS AND HISTAMINOLYTICS

Drugs which together with histamine have affinity and intrinsic activity on histamine receptors may be called histaminomimetics, whereas those which only have affinity to histamine-receptors may be called histaminolytics. Histaminolytics are competitive antagonists of histamine. Therefore many antihistamine drugs, are histaminolytics, but this is not necessarily so for all of them. At present it is not known which moieties in the molecule are essential for intrinsic activity on histamine receptors although the free amino-groups seem to have some significance. When in 2-

TABLE XVII
HISTAMINO-MIMETICS AND -LYTICS

formula	drug	guinea-pig intestine		
		α	pD ₂	pA ₂
	histamine	1	6.5	—
	pyrazole-ethylamine	1	4.1	—
	R = H (pyrethamine)	1	5.4	—
	-C	1	5.3	—
	-C-C	0.9	4.3	—
	-C-C-C	0.1	—	4.0
	-C-C-C-C	0	—	4.2
	R = -C-C-C	0	—	6.0

Ariëns and Simonis, 1960.
Schild, 1957.

pyridylethylamine heavier substituents are introduced in the free amino-group the intrinsic activity gradually decreases so that finally histaminolytics or antihistamines are obtained (see Table XVII) (Ariëns and Simonis, 1960). Consequently the affinity also decreases so that some of the intermediate compounds have such a low affinity for the specific receptors that they behave as non-competitive antagonists. The affinity of the lytics so obtained can be increased by introduction of larger groups of planar ring systems, so apparently, for the lytics, regions adjacent to the receptor for histamine are important for the antihistamines. This suggests that the moieties in histaminolytics, which resemble histamine, are of minor

significance for receptor occupation and that additional moieties mainly determine the affinity. It is then conceivable that the antihistamine drugs and for instance, the atropine-like drugs, have more in common than their respective mimetics.

SYMPATHOMIMETIC DRUGS

According to Ahlquist (1959) a distinction can be made in sympathomimetic drugs between those predominantly acting on α -receptors and those acting on β -receptors. There exist α -sympathomimetics which have both affinity for α -receptors and an α -intrinsic activity, and " α -sympatholytics" which are competitive antagonists of α -mimetics. Furthermore there are β -sympathomimetics which have both β -intrinsic activity

TABLE XVIII
HOMOLOGOUS SERIES OF SYMPATHETIC DRUGS ON α - AND β -RECEPTORS

HO--C-C-N-R (\pm)-compounds

R	α -receptors (<i>vas deferens</i>)			β -receptors (tracheal muscle)		
	α	pD ₂	pA ₂	α	pD ₂	pA ₂
-H	1	5.4	—	1	5.8	—
-C	1	5.7	—	1	6.7	—
	0.4	3.4	—	1	7.5	—
	—	—	< 2.5	1	7.6	—
	0	—	5.5	1	8.1	—
	0	—	5.2	1	8.2	—

DCI	Nethalide

DCI	—	—	—	0	—	4.9
Nethalide	—	—	—	0	—	5.8

Ariens, 1960.
Ariens and Simonis, 1960.
Ariens, Simonis, Waelen and Sonnevile, 1963.

and affinity to β -receptors and " β -sympatholytics." The moieties for the two affinities and two intrinsic activities may be quite different. It should therefore be realised that in SAR studies of sympathomimetics, relative potencies may change and that one type of drug may gradually change into another. In this respect the α : β -affinity ratios and both intrinsic activities are of importance.

SAR studies of derivatives of (\pm)-noradrenaline are presented in Table XVIII. It may be noted that introduction of large substituents on the

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

nitrogen atom causes the α -intrinsic activity to decrease, whereas this operation does not influence the β -intrinsic activity. Furthermore the α -affinity decreases and with the introduction of planar rings it increases again. The β -affinity increases with the introduction of large groups. This implies that predominantly α -sympathomimetics (noradrenaline) change into β -sympathomimetics while the latter compounds in higher doses exert ' α -sympatholytic' properties. The nitrogen group is especially important for α -intrinsic activity whereas the catechol moiety is determinant for β -intrinsic activity. Therefore dichloroisoprenaline (DCI) and especially nethalide (Black and Stephenson, 1962) are β -sympatholytics (see Table XVIII).

In SAR studies it is advantageous to be able to study one particular type of action at a time. This may be achieved by investigating sympathomimetic drugs on the isolated vas deferens that responds only to α -sympathomimetics and isolated tracheal muscle of the calf which is suitable for β -sympathomimetics.

Both α - and β -sympathetic receptors are rather inflexible since there is a substantial difference in the potency of enantiomorphs. The OH group at the C(1) atom seems to be in a key position for both α - and β -receptors. For the β -receptors the difference in potency between the enantiomorphs is about 10–1000, whereas for the α -receptors between 3–10. This would imply that for the β -receptors the position of this OH-group is critical (Ariëns, 1962). It would be very important to resolve the optical isomers of the β -blocking drug, nethalide, to see whether the same configuration is essential for mimetic and blocking action. If the β -mimetic and the β -blocking combine with the same receptor, a great difference in affinity of enantiomorphs of nethalide would be anticipated. The unequivocal notation of configuration is essential for studies of configuration and sympathomimetic action. The use of *d* and *l* and *D* and *L* for rotation and configuration is highly confusing. The use of the sequence rule is pertinent.

It has been pointed out by Burn and Rand (1958), that sympathetic action can be brought about by drugs which do not react with the specific sympathetic receptors but which act by releasing endogenous sympathomimetics from storage sites. This is so with the closely related drugs amphetamine and tyramine. This further complicates the picture. It must be emphasised that in SAR studies these facts should be thoroughly realised.

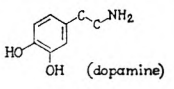
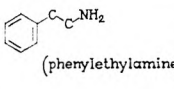
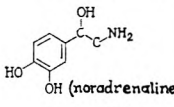
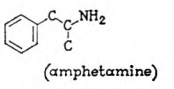
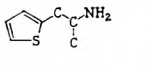
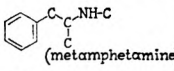
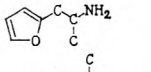
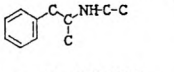
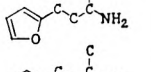
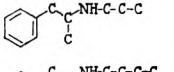
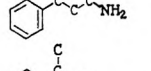
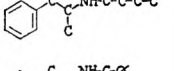
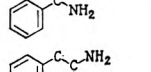
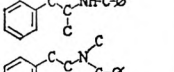
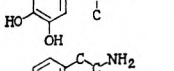
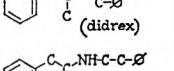
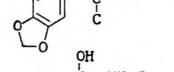
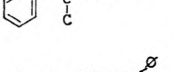
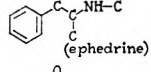
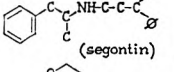
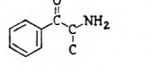
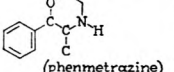
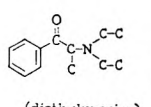
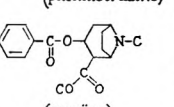
PSYCHOMOTOR STIMULANT DRUGS

Typical psychomotor stimulant drugs are amphetamine, ephedrine and their derivatives. These compounds resemble the catecholamines, noradrenaline, and dopamine in the molecular structure. The relationship between structure and activity of many amphetamine derivatives has been examined by van der Schoot (1961) and van der Schoot and others (1962). Evidence from these experiments is presented in Table XIX. It is essential for psychomotor stimulant action that the drugs be stable and that they be sufficiently lipophilic to pass the blood brain barrier. The

J. M. VAN ROSSUM

TABLE XIX

SAR OF PSYCHOMOTOR STIMULANT DRUGS MEASURED BY INCREASE OF LOCOMOTOR ACTIVITY IN MICE

drug	rel. potency	drug	rel. potency
 (dopamine)	0	 (phenylethylamine)	1-2
 (noradrenaline)	0	 (amphetamine)	100
 (metamphetamine)	100		150-200
 (amphetamine derivative)	35		60
 (amphetamine derivative)	0		50
 (amphetamine derivative)	0		15
 (amphetamine derivative)	0		8
 (didrex)	0		10
 (amphetamine derivative)	15		6
 (ephedrine)	8		0*
 (amphetamine derivative)	50	 (phenmetrazine)	14
 (diethylpropion)	15	 (cocaine)	25

* antagonist
van der Schoot, 1961.

potent drugs therefore have a substituent (e.g. methyl group) on the carbon atom next to the amino-group and they lack phenolic OH-groups.

Furthermore, considering the strict structure activity relationship this may imply that the various amphetamine-like compounds interact with common receptors. There are however some compounds which cause identical psychomotor stimulant actions to amphetamine but which have a completely different structure. One such drug is cocaine. Furthermore the *N*-benzyl derivatives of amphetamine, as for instance, the

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

compound (+)-*N*-benzyl-*N*-methylamphetamine (Didrex), do not show peripheral sympathomimetic actions like amphetamine but do show identical central effects. This may suggest that at least two classes of psychomotor stimulant drugs may exist as was anticipated by van der Schoot (1961).

It has been shown that psychomotor stimulant action is related to the function of catecholamines in the brain whereas the action of 5-HT is not involved (van Rossum, van der Schoot and Hurkmans, 1962; van der Schoot, 1961; van der Schoot and others, 1962; Smith and Dews, 1962). The receptors for catecholamines may be related either to the α -receptors of the peripheral nervous system or to the β -receptors. Since in most amphetamine-like drugs the β -hydroxy group is lacking, a resemblance to β -receptors is unlikely. This could further be established by using selective α - and " β -sympatholytics." The " α -sympatholytics" antagonise psychomotor stimulant action whereas the β -lytics do not influence this effect unless in toxic doses. Now it is likely that receptors for catecholamines in the brain are related to the α -receptors of the peripheral nervous system. We may make a scheme of adrenergic function in the central [nervous

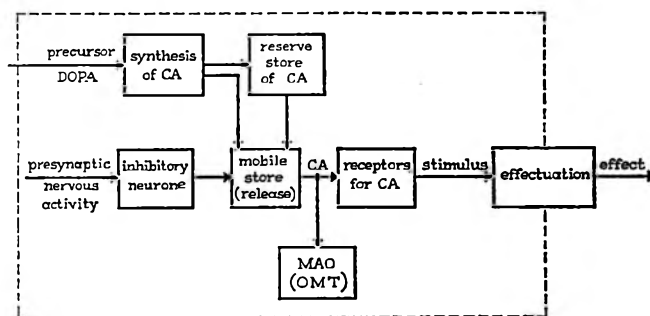


FIG. 6. Scheme of adrenergic function in the brain. The effect is proportional to the concentration of free catecholamines around the receptors. Under normal conditions catecholamines are released by "inhibitory" nervous activity, while catecholamines are continuously synthesised from precursors. Drugs may interfere with adrenergic function at the various blocks for instance by directly reacting with catecholamine receptors or by releasing catecholamines from stores. See text. In this scheme no distinction is made between dopamine and noradrenaline. Obviously in different parts of the brain dopamine and noradrenaline may have a completely different significance.

system (see Fig. 6). Under physiological conditions, catecholamines are synthesised from a precursor DOPA which can pass the blood brain barrier. The catecholamines synthesised are stored in either mobile or reserve stores. Under the influence of the physiological stimuli, catecholamines are released from the mobile stores while this store is refilled from the reserve. Once liberated, catecholamines may occupy specific receptors and elicit an increase in locomotor activity and other effects by a complicated route. The free catecholamines are also subject to metabolism by monoamine oxidase and by conjugation to 3-methoxy derivatives. Consideration of this scheme results in the following list of ten possible

mechanisms of psychomotor stimulant action: (i) drugs that act as precursors of catecholamines (CA); (ii) drugs that inhibit one or more steps in the synthesis of CA like α -methylDOPA; (iii) drugs that inhibit storage of CA; either in the mobile or reserve stores like reserpine; (iv) drugs that stimulate or inhibit the inhibitory neurones and so facilitate or inhibit release of CA from their stores; (v) drugs that cause a release of CA by acting on the stores; (vi) drugs that inhibit the release of CA by acting on the stores; (vii) drugs that inhibit monoamine oxidase or other mechanisms of destruction of CA; (viii) drugs that mimic CA by directly acting on specific receptors; (ix) drugs that antagonise CA by directly acting on specific receptors; (x) drugs that in some way facilitate or inhibit effectuation.

From the point of view of these possibilities, the drugs of immediate interest are those that have a *direct* sympathomimetic action (viii) and those that cause a rise in free CA and thus have an *indirect* sympathomimetic action (iv; v; vii).

Psychomotor stimulants may cause an increase in locomotion by directly reacting with catecholamine receptors or by causing a release of catecholamines from stores and thus causing the concentrations of catecholamines to rise. By pre-treatment with reserpine, catecholamine stores can be depleted so that drugs which act by releasing catecholamines would then become inactive. From experiments like these it could be shown that amphetamine has a direct mechanism of action, that is, amphetamine acts directly on catecholamine receptors, whereas cocaine has an indirect mechanism of action, that is, by releasing catecholamines from stores in the brain (van Rossum, van der Schoot and Hurkmans, 1962; van der Schoot, 1961). A further argument for the indirect action of cocaine is the following experiment: animals pre-treated with reserpine, so that catecholamine stores become depleted and cocaine becomes ineffective, are loaded with DOPA after which cocaine is again injected. DOPA is converted into catecholamines so that stores can be refilled when cocaine is active once more (van Rossum and others, 1962). The time between DOPA injection and cocaine restoration is less than an hour. This might suggest that dopamine instead of noradrenaline is involved in locomotion, since according to biochemical work (Hess, Connamacher, Ozaki and Udenfriend, 1961) the formation of dopamine from DOPA is a fast process, whereas the conversion of dopamine into noradrenaline is very slow. According to the biochemical findings, after one hour there cannot be enough noradrenaline in the brain. Furthermore, Eggels (1963) could not determine any significant amount of noradrenaline at the time that the cocaine effect was restored. At the moment experiments are being made to determine the dopamine concentration at the time of cocaine restoration. A further argument for the involvement of dopamine instead of noradrenaline is the failure to show tachyphylaxis with cocaine. When cocaine is administered at regular intervals, catecholamines are released from stores so that tachyphylaxis was anticipated. But the catecholamine stores can be refilled quickly with dopamine since there is a rapid synthesis from DOPA. The synthesis of noradrenaline from

CHEMICAL STRUCTURE AND BIOLOGICAL ACTIVITY

dopamine, as shown by Hess and others (1961), is a slow process. For the peripheral nervous system it is more likely that noradrenaline is involved so that at that level tachyphylaxis may easily be observed after administration of indirect sympathomimetics.

REFERENCES

- Ahlquist, R. P. (1959). *Pharmacol. Rev.*, **11**, 441-442.
- Ariëns, E. J. (1954). *Arch. int. Pharmacodyn.*, **99**, 32-49.
- Ariëns, E. J., Simonis, A. M. and de Groot, W. M. (1955). *Arch. int. Pharmacodyn.*, **100**, 298-322.
- Ariëns, E. J., van Rossum, J. M. and Simonis, A. M. (1956a). *Arzneimitt.-Forsch.*, **6**, 282-393.
- Ariëns, E. J., van Rossum, J. M. and Simonis, A. M. (1956b). *Ibid.*, **6**, 611-621.
- Ariëns, E. J., van Rossum, J. M. and Simonis, A. M. (1956c). *Ibid.*, **6**, 737-746.
- Ariëns, E. J. and Van Rossum, J. M. (1957a). *Arch. int. Pharmacodyn.*, **110**, 275-299.
- Ariëns, E. J. and van Rossum, J. M. (1957b). *Ibid.*, **113**, 89-100.
- Ariëns, E. J. (1960). *Ciba Foundation Symposium on Adrenergic Mechanisms*, Ed. Wolstenholme, p. 264, London: J. and A. Churchill Ltd.
- Ariëns, E. J., van Rossum, J. M. and Koopman, P. C. (1960). *Arch. int. Pharmacodyn.*, **127**, 459-478.
- Ariëns, E. J. and Simonis, A. M. (1960). *Ibid.*, **127**, 479-495.
- Ariëns, E. J. and Simonis, A. M. (1962). *Acta Physiol. Pharmacol., Neerl.*, **11**, 151-172.
- Ariëns, E. J. and Simonis, A. M. (1963). *Acta Physiol. Pharmacol. Neerl.*, in the press.
- Ariëns, E. J. (1962). *Symposium in Modern Concepts in Relationships Between Structure and Biological Activity*, Vol. 7, 247-265. Proc. Ist. Intern. Pharmacol. meeting, Stockholm 1961. Oxford: Pergamon Press.
- Ariëns, E. J., Simonis, A. M. Waelen, M. J. G. A. and Sonnevile, P. F. (1963). *Arch. exp. Path. Pharmacol.*, in the press.
- Barlow, R. B. (1960). *Steric Aspects of Drug Action.*, Biochem. Soc. Symp. No. 19, 44-67.
- Beckett, A. H. (1962). In *Proc. Int. Union Physiol. Sci.*, XXII Intern. Congress I, p. 805. Leiden.
- Beckett, A. H. (1959). *Stereochemical Factors in Biological Activity*. In *Fortschritte der Arzneimittelforschung*, Ed. Jucker, Ernst I, p. 455, Basle: Birkhäuser Verlag.
- Black, J. W. and Stephenson, J. S. (1962). *Lancet*, **2**, 311-314.
- Burn, J. H. and Rand, M. J. (1958). *J. Physiol.*, **144**, 314-336.
- Cahn, R. S. and Ingold, C. K. (1951). *J. chem. Soc.*, 612-622.
- Cahn, R. S., Ingold, C. K. and Prelog, V. (1956). *Experientia*, **12**, 81-95.
- Carlsson, A., Lindqvist, M. and Magnusson, T. (1957). *Nature, Lond.*, **180**, 1200.
- Chiavarelli, S. and Setting, G. (1958). *Gaz. Chim. Ital.*, **88**, 1234-1241.
- Dirkx, I. P. (1962). Ph.D. Thesis, Municipal University, Amsterdam.
- Eccles, J. C. (1961). In *K. Kramer Ergebn. der Physiol.*, 51E Bd, p. 370, Berlin: Springer Verlag.
- Eggs, P. (1963). Ph.D. Thesis, State University of Utrecht, in the press.
- Ellenbroek, B. W. J. and van Rossum, J. M. (1960). *Arch. int. Pharmacodyn.*, **125**, 216-220.
- Ellenbroek, B. W. J. (1963). Ph.D. Thesis, Cath. University of Nijmegen, in the press.
- Ferguson, J. (1939). *Proc. Roy. Soc., Lond.*, **B**, **127**, 387-405.
- Freudenberg, E. (1932). *Stereochemie*, p. 697, Leipzig: Deuticke.
- Furchgott, R. F. (1955). *Pharmacol. Rev.*, **7**, 183-245.
- Gaddum, J. H., Hameed, K. A., Hathway, D. E. and Stephens, F. J. (1955). *Quat. J. exp. Physiol.*, **40**, 49-75.
- Gyermek, L. and Unna, K. R. (1960). *J. Pharmacol.*, **128**, 37-40.
- Hanna, C. (1960). *Arch. int. Pharmacodyn.*, **129**, 191-201.
- Harms, A. F. (1956). Ph.D. Thesis, Free University, Amsterdam.
- Hess, S. M., Connamacher, R. H., Ozaki, M. and Udenfriend, S. (1961). *J. Pharmacol.*, **134**, 129-148.
- Jaramillo, J. (1962). M.S. Thesis, Tulane University, New Orleans.
- Koopman, P. C. (1960). Ph.D. Thesis, Cath. University, Nijmegen, Rotterdam: Bronder Offset.

J. M. VAN ROSSUM

- Kroneberg, G. (1955). *Arch. exp. Path. Pharmacol.*, **225**, 522-532.
- Laarhoven, W. M., Nivard, R. J. F. and Havinga, E. (1961). *Experientia*, **17**, 214-215.
- Long, J. P., Luduena, F. P., Tullar, B. F. and Lands, A. M. (1956). *J. Pharmacol.*, **117**, 29-38.
- Longo, V. G. and Chiavarelli, S. (1962). *Arch. int. Pharmacodyn.*, **138**, in press.
- Luduena, F. P. (1962). *Ibid.*, **137**, 155-166.
- Luduena, F. P., von Euler, L., Tullar, B. F. and Lands, A. M. (1957). *Ibid.*, **91**, 392-401.
- Lyle, G. G. (1960). *J. org. chem.*, **25**, 1779-1784.
- Nickerson, M. (1956). *Nature, Lond.*, **178**, 697-698.
- Pauling, L. (1961). *Science*, **134**, 15-21.
- Pfeiffer, C. C. (1956). *Science*, **124**, 29-31.
- Pratesi, P., La Manna, A., Campiglio, A. and Ghislandi, V. (1958). *J. chem. Soc.*, 2069-2074.
- Pratesi, P., La Manna, A., Campiglio, A. and Ghislandi, V. (1959). *Ibid.*, 4062-4065.
- van Dijk, J. and Moed, H. D. (1959). *Recueil Trav. Chim., Pays Bas*, **78**, 22-30.
- van Dijk, J. and Moed, H. D. (1961). *Ibid.*, **80**, 574-588.
- van Rossum, J. M. (1958). Ph.D. Thesis, University, Nijmegen, Bruges: St. Catherine Press.
- van Rossum, J. M., Ariëns, E. J. and Linssen, G. H. (1958). *Biochem. Pharmacol.*, **1**, 193-199.
- van Rossum, J. M. and Ariëns, E. J. (1959a). *Arch. int. Pharmacodyn.*, **118**, 393-417.
- van Rossum, J. M. and Ariëns, E. J. (1959b). *Ibid.*, **118**, 418-446.
- van Rossum, J. M. and Ariëns, E. J. (1959c). *Ibid.*, **118**, 447-466.
- van Rossum, J. M. (1960a). *Science*, **132**, 954-956.
- van Rossum, J. M. (1960b). *Experientia*, **16**, 373-375.
- van Rossum, J. M. and E. J. Ariëns (1962). *Arch. int. Pharmacodyn.*, **136**, 385-413.
- van Rossum, J. M., van der Schoot, J. B. and Hurkmans, J. A. Th.M. (1962). *Experientia*, **18**, 229-230.
- van Rossum, J. M. and Hurkmans, J. A. Th.M. (1962). *Acta Physiol. Pharmacol. Neerl.*, **11**, 173-194.
- van Rossum, J. M. (1962). *Int. J. Neuropharmacol.*, **1**, 97-110; 403-421.
- Roth, F. E. and Govier, W. M. (1958). *J. Pharmacol.*, **124**, 347-348.
- Schild, H. O. (1947). *Brit. J. Pharmacol.*, **2**, 251-258.
- Schild, H. O. (1949). *Ibid.*, **4**, 277-280.
- Schild, H. O. (1957). *Pharmacol. Rev.*, **9**, 242-246.
- Schoot, J. B. van der (1961). Ph.D. Thesis, University, Nijmegen, Nijmegen: Thoben Offset.
- Schoot, J. B. van der, Ariëns, E. J., van Rossum, J. M. and Hurkmans, J. A., Th.M. (1962). *Arzneimitt.-Forsch.*, **12**, 902-907.
- Smith, C. B. and Dews, P. B. (1962). *Psychopharmacol.*, **3**, 55-59.
- Stelt, C. van der, Harms, A. F. and Nauta W. Th. (1961). *J. med. Pharm. Chem.*, **4**, 335-349.
- Stephenson, R. P. (1956). *Brit. J. Pharmacol.*, **11**, 379-393.
- Waser, P. G. (1958). *Experientia*, **14**, 356-359.
- Waser, P. G. (1961). *Pharmacol. Rev.*, **13**, 465-515.

RESEARCH PAPERS

THE CRITICAL MICELLE CONCENTRATIONS OF DOUBLE LONG-CHAIN ELECTROLYTES (AMINE SOAPS) IN AQUEOUS SOLUTION

BY A. PACKTER AND M. DONBROW

From the School of Pharmacy, Chelsea College of Science and Technology, Manresa Road, London, S.W.3

Received December 17, 1962

Micelle formation and gegenion binding have been studied in aqueous solutions of primary and quaternary ammonium dodecyl sulphates (amine soaps) by the conductivity method. Critical micelle concentrations decrease markedly with increase in the cation chain length; this phenomenon is due mainly to increase in the free energy of micelle formation. Gegenion binding also increases with increasing cation chain length and reaches 95 per cent for the octyl and higher ammonium salts.

DOUBLE long-chain electrolytes (amine soaps), in which both anion and cation are attached to long-chain organic groups are used as surface-active agents when good solubility is required in both water and organic phases; the hydrophilic-lipophilic balance is often modified advantageously when organic counterions are substituted in place of the more usual inorganic ions in such materials (Moilliet, Collie and Black, 1961; Clayton and Sumner, 1954; Winsor, 1954).

Organic counterions are also known to have a marked influence on micelle formation by colloidal electrolytes in solution; the critical micelle concentration (CMC) decreases markedly with increase in counterion length (Cushman, Brady and McBain, 1948; Goddard, Harva and Jones, 1953; Scott and Tartar, 1943). This phenomenon might also be advantageous in many practical applications (Clayton and Sumner, 1954; Winsor, 1954) and in this respect, amine soaps may constitute a new and distinctive class of surface-active agents.

Few detailed studies on the physical properties of these materials have been reported and a systematic investigation of micelle formation and allied phenomena in solution should be of theoretical and practical interest.

In this initial work, the CMC of a series of alkyl ammonium dodecyl sulphates have been determined by the conductivity method.

The effect of counterion size on micelle formation and counterion binding has also been studied.

EXPERIMENTAL

Materials

The "amine soap" solutions studied were prepared *in situ* by exact neutralisation of a prepared batch of aqueous dodecyl sulphuric acid solution (0.1M) with various organic bases.

Sodium dodecyl sulphate. "Sipons" W.D. product was used. This material contained less than 0.1 per cent free long-chain alcohols.

The sulphate was hydrolysed to "lauryl alcohol" and the latter analysed by gas chromatography (on a Celite—20 per cent Silicone E301 column) and shown to be a mixture of approximately 92 per cent dodecyl and 8 per cent tetradecyl alcohols.

Dodecyl sulphuric acid was prepared by passing a solution (0.1 M) of the sodium dodecyl sulphate through a column of "Zeo-Karb 215" (H).

The acid solutions were standardised by potentiometric titration with sodium hydroxide. Complete conversion to dodecyl sulphuric acid was checked by determination of the equivalent conductivity of the neutralised acid; unchanged sodium dodecyl sulphate would lead to high equivalent conductivity values.

Primary amines. Methylamine and ethylamine of above 99 per cent purity were supplied by British Drug Houses Ltd., redistilled butylamine and hexylamine of 98–99 per cent purity and octylamine of 95–98 per cent purity by L. Light and Co., Ltd.

Purity was checked by equivalent weight determination and boiling-point (or melting-point).

Tetramethyl ammonium bromide was supplied by British Drug Houses Ltd., and recrystallised from alcohol.

Alkyl trimethylammonium bromides were prepared by reacting the alkyl bromide with excess alcoholic trimethylamine (Scott and Tartar, 1943).

Ethyl and butyl bromide (of above 99 per cent purity) were supplied by British Drug Houses Ltd. Hexyl, octyl and decyl bromide (of 95–96 per cent purity) were supplied by L. Light and Co., Ltd. Their purity was checked by gas chromatographic analysis on a Celite—10 per cent tricresyl phosphate column. These bromides contained up to 3 per cent isomers, about 0.5 per cent of lower homologues and about 2 per cent of higher homologues; this distribution should have little effect on CMC values (Harkins, 1953).

Alcohol and excess trimethylamine were removed by evaporation on a water bath. The quaternary bromides were then recrystallised from alcohol or acetone.

Quaternary ammonium hydroxide solutions were prepared by treating the corresponding halide salt solution (0.1 M) with a slight excess of silver oxide; the solution was then centrifuged and filtered free of silver halide. Complete conversion to hydroxide was checked by testing with acidified silver nitrate solution and by potentiometric titration of the prepared hydroxide with standard hydrochloric acid.

Alkyl ammonium dodecyl sulphates. Dodecyl sulphuric acid solutions (0.1 M) were standardised by potentiometric titration with sodium hydroxide.

These solutions were then titrated potentiometrically to pH 7.0* with the appropriate primary or quaternary ammonium hydroxide solution and the salt solutions were diluted as required.

* In the case of the primary amine soaps, a slight excess of free hydroxide was present in the titrated solution to prevent conductivity errors caused by hydrolysis at the higher dilutions.

CMC OF DOUBLE LONG-CHAIN ELECTROLYTES

The water used for the preparation of solutions and for conductivity measurements was prepared from freshly glass-distilled water (specific conductivity of $3-5 \times 10^{-6}$ ohms $^{-1}$) by passing through a bed of mixed ion-exchange resins (in an "Elgostat") and used immediately.

Its specific conductivity, at 25°, varied between 0.8 and 1.2×10^{-6} ohms $^{-1}$.

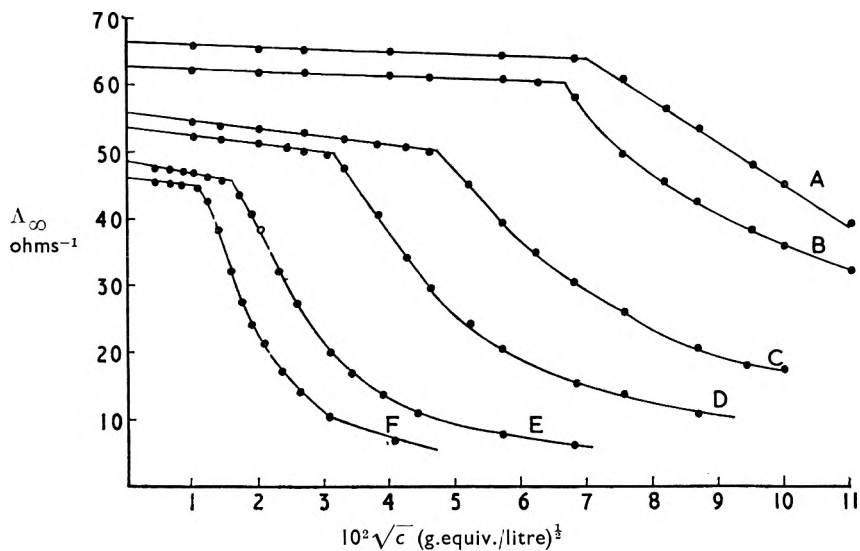


FIG. 1. Variation of equivalent conductivity of alkyl ammonium dodecyl sulphates with concentration (25°).

A, Tetramethyl; B, Ethyl trimethyl; C, Butyl trimethyl; D, Hexyl trimethyl; E, Octyl trimethyl; F, Decyl trimethyl.

Measurement of Critical Micelle Concentrations and Allied Properties

Critical micelle concentrations were measured by the conductivity method (Scott and Tartar, 1943; Evans, 1956).

A titration cell was used, with platinised platinum dipping electrodes, similar to that described by previous workers (Winsor, 1950; Evans, 1956); the cell constant was determined using standard potassium chloride solutions. Solutions were stirred with flowing nitrogen.

Conductivity measurements were made, at $25.0 \pm 0.1^\circ$, using a "Pye" Conductance Bridge (Model 11700) operating at 300 c./sec. and fitted with a cathode ray oscillographic detector; cell capacitance effects were balanced out by means of a variable condenser in parallel with the variable resistance. The Bridge operated with an accuracy of 0.1 per cent.

Measured volumes of the alkyl ammonium dodecyl sulphate solution were run into conductivity water, the solutions allowed to come to equilibrium at 25°, stirring stopped and the conductivity measured.

In all, 15 to 20 measurements were taken with each salt to provide a suitable range of conductivities for studies both below and above the CMC; generally, the range 10^{-4} N to $5 \times$ CMC was investigated.

RESULTS AND DISCUSSION

Determination of Critical Micelle Concentrations

Typical *equivalent conductivity—(concentration)^{1/2}* graphs for a series of alkyl ammonium dodecyl sulphate solutions studied are presented in Fig. 1. The Λ_{∞} values decrease regularly with increasing chain length and decreasing cation mobility.

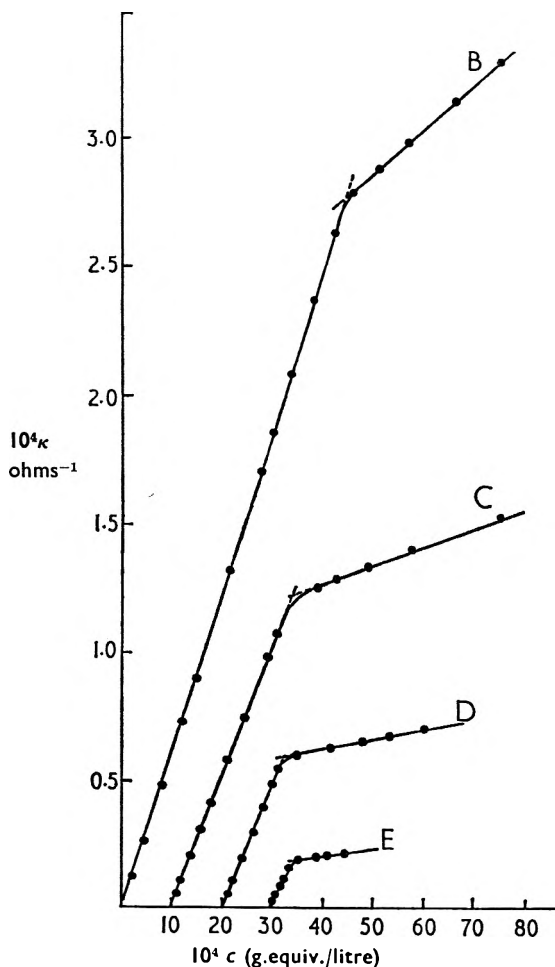


FIG. 2. Variation of specific conductivity of alkyl ammonium dodecyl sulphates with concentration (25°). For key, see Fig. 1. The origin has been shifted a scale division dexterwise for the concentration values of the butyl, hexyl and octyl trimethylammonium salts, to ensure greater clarity.

Formation of micelles in dilute aqueous solutions of these salts is demonstrated by the sharp breaks in these graphs (Scott and Tartar, 1943; Evans, 1956). The critical micelle concentration is seen to decrease markedly with increase in length of the cation chain.

CMC OF DOUBLE LONG-CHAIN ELECTROLYTES

Specific conductivity—concentration graphs were also plotted. The results presented in Fig. 2 are typical of those obtained for all the systems studied.

Below the CMC the slopes (S_1) decrease with increasing cation chain length and decreasing cation mobility. Above the critical range, the gegenion-micelle interaction is increasing with increase in cation chain length (Evans, 1956) and the slopes (S_2) decrease even more sharply.

Critical micelle concentrations were determined from the intersection of the two linear portions of the specific conductivity graphs.

The results are summarised in Table I. The limits of error were ± 0.02 mM for all CMC values.

TABLE I
CRITICAL MICELLE CONCENTRATION, OF ALKYL AMMONIUM DODECYL SULPHATES (25°)

	CMC (mM)	$-\frac{\Delta G^\circ}{RT}$
Tetramethylammonium	5.50	16.8
Ethyl trimethylammonium	4.30	17.5
n-Butyl trimethylammonium	2.38	18.9
n-Hexyl trimethylammonium	1.25	20.5
n-Octyl trimethylammonium	0.40	23.0
n-Decyl trimethylammonium	0.21	24.3
Ammonium	6.16	16.6
Methylamine	5.70	17.0
Ethylamine	5.00	17.3
n-Butylamine	2.92	18.5
n-Hexylamine	1.12	20.9
n-Octylamine	0.28	23.7
n-Decylamine	sparingly soluble	

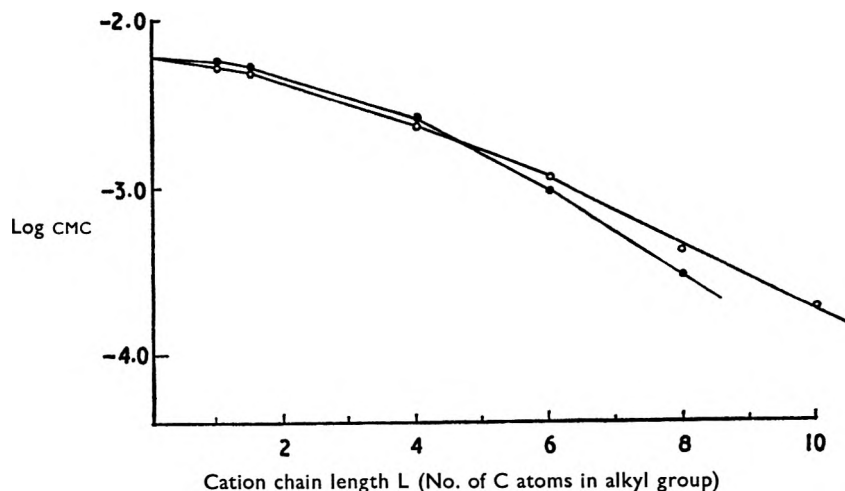


FIG. 3. Effect of cation chain length on critical micelle concentrations (25°).

The CMCs decrease markedly with increasing cation chain length. The hexyl- and octylamine salts have lower CMC values than the corresponding quaternary trimethyl ammonium salts. Log CMC values are plotted against chain length (L) in Fig. 3.

Micelle Formation and Structure

Phillips (1955) has proposed the following idealised equilibrium for micelle formation with long-chain electrolytes,



where X^- represents the dodecyl sulphate anion,

R^+ represents the smaller gegenion,

$[X_N R_{N-p}]^{p-}$ represents the micelle,

N represents the number of anions in the micelle,

$(N-p)$ represents the number of the smaller cations in the micelle,

p represents the net effective charge on the micelle.

The size and molecular weight of the micelle will be determined by the value of $(2N-p)$; while the ratio $(N-p)/N$ represents the fraction of gegenions bound on the surface or within the micelle or both. Phillips (1955), Stigter (1954) and Hoyer (1957) have shown that CMC values are hardly affected by micelle size but depend on the p/N ratio. In the absence of added electrolyte, CMC values are mainly determined by $-\Delta G^\circ$, the standard free energy decrease associated with the formation of one g. mole. of micelle from its constituent ions. Phillips (1955) has proposed the relation,

$$\log_{10} \text{CMC} = \frac{-\Delta G^\circ}{2.303 (2-p/N)RT} \quad \dots \quad (2)$$

Counterion Binding

Integral values of p and $(N-p)/N$ for the systems studied were estimated from the conductivity data, using Evans' (1956) equation.

$$\frac{p^2}{N^{4/3}} (1,000 S_1 - \lambda_{R^+}) + \frac{p\lambda_{R^+}}{N} = 1,000 S_2 \quad \dots \quad (3)$$

where S_1 is the gradient of the specific conductivity-concentration graph below the CMC, S_2 is the gradient above the CMC, λ_{R^+} is the cation mobility.

Since p and $(N-p)/N$ values are practically unaffected by large changes in N (Evans, 1956), e.g. from 40 to 120, Phillips' value of $N = 80$ for sodium dodecyl sulphate was used in all the calculations.

The values calculated for the alkyl ammonium dodecyl sulphate micelles studied are summarised in Table II: data for the ammonium salt are also included.

The micelle charge (p/N) decreases from 0.23 for the ammonium salt to 0.05 for the higher alkyl ammonium dodecyl sulphates; that is, the degree of binding $(N-p)/N$ of the organic gegenions increases from 77 to 95 per cent.

Energetics of Micelle Formation

The increase in $(N-p)/N$ ratios is insufficient to explain the marked reduction in CMC values; the latter effect must therefore be mainly attributed to a significant increase in the free energy change (ΔG°) of micelle formation.

CMC OF DOUBLE LONG-CHAIN ELECTROLYTES

The standard free energy decrease (ΔG°) of micelle formation in the systems studied was estimated from the CMC results using equation 2 (Phillips, 1955),

$$-\Delta G^\circ = 2.303 (2-p/N)RT \log_{10} \text{ CMC calories per g. mole.}$$

where CMC values are expressed in *mole fractions*.

The results are summarised in Table I.

The free energy change increases progressively with length of the organic cation. The estimated ($-\Delta G^\circ$) values for the alkyl ammonium salts are from 1 to 7RT calories per g. mole. greater than the value for the ammonium dodecyl sulphate and it is evident that organic cation—organic anion interactions must contribute to the overall free energy of micelle formation.

TABLE II

DEGREE OF CATION BINDING IN ALKYL AMMONIUM DODECYL SULPHATES ESTIMATED FROM SPECIFIC CONDUCTIVITY DATA AT 25° (CALCULATED FOR N = 80)

Salt	λ_{R+}	1,000 S ₁	1,000 S ₂	p	$\frac{(N-p)}{N}$
Ammonium	73.4	91.2	30.0	17.5	0.77
Tetramethylammonium	44.9	63.8	19.0	14.1	0.82
Ethyl trimethylammonium	40.5	57.0	17.0	13.3	0.83
n-Butyl trimethylammonium	33.3	50.0	9.2	9.9	0.88
n-Hexyl trimethylammonium	29.2	47.0	4.2	6.1	0.92
n-Octyl trimethylammonium	26.2	44.5	3.2	5.1	0.94
n-Dodecyl trimethylammonium	24.0	43.0	2.5	4.9	0.94
Methylamine	57.8	75.4	16.0	12.1	0.85
Ethylamine	44.9	65.0	15.0	11.3	0.86
n-Butylamine	35.5	55.5	9.5	9.6	0.88
n-Hexylamine	32.5	53.0	4.4	5.9	0.93
n-Octylamine	30.5	50.0	2.6	4.8	0.94

This could take place either as a result of changes in electrostatic effects at the anion heads or by enhancement of van der Waals' interactions within the micelle.

The electrostatic repulsion between adjacent anion heads is indeed not very marked, even in the case of sodium dodecyl sulphate micelles and reduction of this repulsion by ion-pair formation with organic cations would lead at most to an increase in ($-\Delta G^\circ$) of 0.6 RT calories per g. mole (Phillips, 1955). However, the large increase in free energy change observed could be explained by a closer packing of the organic sections of the long-chain anions inside the micelle, which might occur as a result of the reduction of repulsive forces at the anion heads. Such closer packing would in turn lead to an increase both in ($-\Delta H$), the heat of interaction of the hydrocarbon chains and in ($-T\Delta S$), the entropy of micelle formation (Pethica, 1958).

On the other hand, Goddard and others (1953) have suggested that in aqueous solutions of double long-chain electrolytes, the organic cations also enter the micelles to form a palisade structure with the organic anions. This penetration would lead to van der Waals' interaction between the organic sections of cation and anion, similar to that observed on addition of long-chain alcohols to aqueous solutions of ordinary colloidal electro-

A. PACKTER AND M. DONBROW

lytes. Such interaction would increase with increasing cation chain length and would enhance the usual interaction between the organic sections of the anions (Ooshika, 1954).

REFERENCES

- Clayton, W. and Sumner, C. G. (1954). *Emulsions*, p. 165, 526, London: J. and A. Churchill.
- Cushman, A., Brady, A. P. and McBain, J. W. (1948). *J. Colloid Sci.*, **3**, 425-435.
- Evans, H. C. (1956). *J. chem. Soc.*, 579-586.
- Goddard, E. D., Harva, O. and Jones, T. G. (1953). *Trans. Faraday Soc.*, **49**, 980-984.
- Harkins, W. D. (1953). *The Physical Chemistry of Surface Films*, p. 310, New York: Reinhold.
- Hoyer, W. D. (1957). *J. Phys. Chem.*, **61**, 1283-1285.
- Moilliet, J. L., Collie, B. and Black, W. (1961). *Surface Activity*, p. 492, London: E. and F. Spon Ltd.
- Ooshika, T. (1954). *J. Colloid Sci.*, **9**, 254-262.
- Pethica, B. A. (1958). *Trans. Faraday Soc.*, **54**, 587-593.
- Phillips, J. N. (1955). *Trans. Faraday Soc.*, **51**, 561-569.
- Scott, A. B. and Tartar, H. V. (1943). *J. Amer. chem. Soc.*, **65**, 692-698.
- Stigter, D. (1954). *Rec. Trav. Chim., Pays Bas*, **73**, 611-625.
- Winsor, P. A. (1950). *Trans. Faraday Soc.*, **46**, 762-772.
- Winsor, P. A. (1954). *Solvent Properties of Amphiphilic Compounds*, p. 33, 51, 101, London: Butterworths.

THE QUANTITATIVE CONVERSION OF BARBALOIN TO ALOE-EMODIN AND ITS APPLICATION TO THE EVALUATION OF ALOES

BY J. W. FAIRBAIRN AND S. SIMIC

From the Department of Pharmacognosy, School of Pharmacy, University of London, Brunswick Square, London, W.C.1

Received November 19, 1962

Barbaloin can be quantitatively converted into aloe-emodin by heating a solution in 4 *N* hydrochloric acid, containing 4 per cent ferric chloride, for 4 hr. in a boiling water bath under reflux. Each of these numerical conditions can be varied ± 10 per cent without significantly altering the yields. The reaction has been successfully applied to the assay of commercial samples of aloes.

BARBALOIN, 10(1)-deoxyglucosyl aloe-emodin anthrone, is an important constituent of aloes and has recently been shown to occur in cascara (*Rhamnus purshiana* D.C. bark) in the form of glycosides (Fairbairn and Simic, 1960). A method of determining the barbaloin content of these crude drugs would therefore be useful in their evaluation.

Numerous attempts have been made to determine barbaloin in aloes and its preparations. Of published methods (Brody, Voigt and Maher, 1950; Mary, Christiansen and Beal, 1956; Borkowski, Henneberg and Urszulak, 1960; Janiak and Böhmert, 1962; Kraus, 1957; Paris and Durand, 1956; Lister and Pride, 1959) none was considered satisfactory, mainly because of interfering substances in the crude drugs, especially in cascara. The conversion of barbaloin to aloe-emodin offered an alternative since aloe-emodin may be readily separated from impurities and estimated colorimetrically. Various reagents have been described for this conversion and of these we have found ferric chloride the most satisfactory (cf. Cahn and Simonsen, 1932; Harders, 1949; Hay and Haynes, 1956; Betts, 1961; Paris and Durand, 1956; Hörhammer, Wagner and Föcking, 1959). The recommended method is given in the experimental section. The results obtained on standard solutions of barbaloin are given in Table I.

In aloes, small quantities of free anthraquinones and normal glycosides are present; these are removed by the procedure described in the first paragraph in the section on "Estimation of the Barbaloin Content of Aloes".

Several authors have claimed that when extracts of aloe are chromatographed, a spot of R_f value less than barbaloin may be observed (Awe, Auterhoff and Wachsmuth-Melm, 1954; Awe and Kümmell, 1960). Unlike barbaloin, this spot, when treated with alkali, fluoresces blue in ultra-violet light: it has been suggested that the substance responsible is an isomer of barbaloin. Janiak and Böhmert (1962) claim it can be removed from aloe solution using a column of polyamide, but we have found different samples of polyamide to differ in their capacity to achieve this separation. Using paper chromatography, we separated some of

this material which, on treatment with ferric chloride, gave only a faint pink colour with sodium hydroxide in contrast to the deep red colour of the simultaneously eluted barbaloin. The faint colour may well be due to traces of barbaloin incompletely separated from the blue fluorescent substances. It did not interfere in our assay. Table II shows the results of applying our method to 5 commercial samples of curaçao aloes: the results are compared with those obtained by the calcium precipitation method (Lister and Pride, 1959).

EXPERIMENTAL

Materials

Barbaloin, prepared by the method of Hay and Haynes (1956), was crystallised from water then methanol in lemon yellow needles. When dried to constant weight, at 110 to 120° over magnesium perchlorate, it lost 4.6 per cent (mean of 4 analyses). On exposure to air the dried material very rapidly recovered its moisture content; after 15 min. 3.7 per cent gain; after 30 min. 4.4 per cent gain.

Found (anhydrous material) C, 60.5, 60.0; H, 5.3, 5.3. Calculated for $C_{21}H_{22}O_9$; C, 60.3; H, 5.3. Found (before drying): mean of 5 analyses; C, 57.6; H, 5.6. Calculated for $C_{21}H_{22}O_9, H_2O$: C, 57.8; H, 5.5; H_2O , 4.1 per cent. For the anhydrous material, m.p. 148–149° (lit. (Hay and Haynes, 1956) gives m.p. 148–148.5°); λ_{max} 269, 296.5 and 354 $m\mu$; (*E* (1 per cent, 1 cm.) 192, 226 and 259 respectively).

TABLE I

YIELDS OF ALOE-EMODIN (MG./100 MG. BARBALOIN) USING THREE SOLUTIONS OF BARBALOIN. EACH SOLUTION WAS ASSAYED 4 TO 6 TIMES BY THE METHOD DESCRIBED

Solution 1 27.6 mg. barbaloin/litre	Solution 2 27.8 mg. barbaloin/litre	Solution 3 62.4 mg. barbaloin/litre
61.6	63.1	62.4
60.7	62.8	61.4
61.0	62.8	62.0
60.5	62.0	61.0
61.8	Mean 62.7	Mean 61.7
61.0		
Mean 61.1		

Grand Mean = 61.8 (s.d., 0.84).

100 per cent theoretical yield = 61.9 ($100 \times C_{13}H_{19}O_3/C_{21}H_{22}O_9, H_2O$).

Aloe-emodin, prepared by the method of Cahn and Simonsen (1932) was sublimed at 150° and 0.001/mm. Hg and crystallised from glacial acetic acid as orange needles, m.p. 225–226°, λ_{max} (N sodium hydroxide) 500 $m\mu$; [*E* (1 per cent, 1 cm.); 320; at λ 440 $m\mu$; 142].

Conversion of barbaloin to aloe-emodin. The following details represent the optimum conditions* for the hydrolysis. Barbaloin dissolved in 4N hydrochloric acid containing 4 per cent ferric chloride is heated at 100° for 4 hr. in a boiling water-bath under reflux. The cooled solution

* The amount of ferric chloride, HCl and the time can be varied ± 10 per cent without altering the yields.

CONVERSION OF BARBALOIN TO ALOE-EMODIN

was then extracted with carbon tetrachloride[†] and assayed colorimetrically as described in the method. The yield of aloe-emodin from the barbaloin monohydrate (mean of 14 results, Table I) was 61.8 mg./100 mg. (theoretical yield, 61.9 mg.) and from the anhydrous barbaloin (mean of two results) was 63.5 mg./100 mg. (theoretical yield, 64.6 mg.).

Ammonium persulphate (Seel, 1919) and sodium metaperiodate (Hay and Haynes, 1956) gave variable results ranging from 0 to 35 mg./100 mg. barbaloin.

TABLE II
BARBALOIN CONTENT (PER CENT AIR-DRY MATERIAL) OF COMMERCIAL SAMPLES OF CURAÇAO ALOES

Sample	Present method	Ca precipitation method
1	$\left. \begin{array}{c} 30.1 \\ 28.4 \\ 29.4 \end{array} \right\} 29.3$	22.5
2	$\left. \begin{array}{c} 31.5 \\ 31.1 \end{array} \right\} 31.3$	23.5
3	$\left. \begin{array}{c} 32.5 \\ 33.5 \end{array} \right\} 33.0$	22.9
4	$\left. \begin{array}{c} 33.3 \\ 32.8 \end{array} \right\} 33.1$	21.0
5	$\left. \begin{array}{c} 33.5 \\ 33.1 \end{array} \right\} 33.3$	22.6

Estimation of the Barbaloin Content of Aloes

Transfer about 0.2 g. powdered aloe sample, accurately weighed, to a 200 ml. flask, moisten with 2 ml. methanol, add 80 ml. hot water and shake for 30 min. Cool, filter into a 100 ml. volumetric flask and make up to volume. To 10 ml. of this solution add 1 ml. hydrochloric acid B.P. and heat for 15 min. in a boiling water-bath. Cool, extract with 2 × 20 ml. carbon tetrachloride, wash the combined tetrachloride layers with 2 × 10 ml. water, discard the carbon tetrachloride layer and return the washings to the aqueous acid layer; transfer this to a 100 ml. volumetric flask and make up to volume.

To 10 ml. of this solution add 6 ml. hydrochloric acid B.P. and 0.6 g. anhydrous ferric chloride and heat in a boiling water-bath under reflux for 4 hr. Cool, extract with 3 × 20 ml. carbon tetrachloride and wash the combined carbon tetrachloride extracts with 2 × 10 ml. water. Reject the washings. Extract the carbon tetrachloride layer with 15, 5 and 5 ml. of N sodium hydroxide; heat the combined alkaline extracts in a boiling water bath for 5 min. (to drive off traces of carbon tetrachloride), cool and make up to 25 ml. Determine the extinction of this solution, at 500 m μ within 1 hr. and estimate the concentration of aloe-emodin from the *E* (1 per cent, 1 cm.) value of 320 or from a suitable calibration curve. Calculate the percentage of barbaloin present from the fact that 1 mg. aloe-emodin is equivalent to 1.61 mg. C₂₁H₂₂O₉·H₂O.

[†] Carbon tetrachloride is a specific solvent for aloe-emodin and is thus preferable to the ether or butanol used by other workers (Auerhoff and Ball, 1954; Harders, 1949; and cf. Fairbairn, 1942).

The absorption curve of this alkaline solution between 440 and 550 μ was in close agreement with the curve obtained from a sample of pure aloe-emodin in an alkaline solution of the same concentration.

Acknowledgements. We would like to thank Westminster Laboratories Ltd. for a grant in aid of one of us (S.S.), Mr. D. Watt (MacFarlan Smith Ltd., Edinburgh) for the samples of aloes and the analytical figures quoted in the last column of Table II, S. B. Penick and Co., New York, for the sample of aloe-emodin, and Mr. G. S. Crouch, School of Pharmacy, for the microanalyses of our barbaloin samples.

REFERENCES

- Auterhoff, H. and Ball, B. (1954). *Arzneimitt.-Forsch.*, **4**, 725-729.
Awe, W., Auterhoff, H. and Wachsmuth-Melm, C. L. (1958). *ibid.*, **8**, 243-245.
Awe, W. and Kümmell, H. J. (1960). *Arch. Pharm., Berl.*, **293**, 271-277.
Betts, T. J. (1961). Thesis, p. 105, University of London.
Borkowski, B., Henneberg, M. and Urszulak, I. (1960). *Biul. Inst. Rosl. Leczn.*, **6**, 125-135.
Brody, T. M., Voigt, R. F. and Maher, F. T. (1950). *J. Amer. pharm. Ass. Sci. Ed.*, **39**, 666-672.
Cahn, R. S. and Simonsen, J. L. (1932). *J. chem. Soc.*, 2573-2582.
Fairbairn, J. W. (1942). *Pharm. J.*, **148**, 198-199.
Fairbairn, J. W. and Simic, S. (1960). *J. Pharm. Pharmacol.*, **12**, 45T-51T.
Harders, C. L. (1949). *Pharm. Weekbl.*, **84**, 250-258, 273-281.
Hörhammer, L., Wagner, H. and Föcking, O. (1959). *Pharm. Ztg., Berl.*, **104**, 1183-1186.
Hay, J. E. and Haynes, L. J. (1956). *J. chem. Soc.*, 3141-3147.
Janiak, B. and Böhmert, H. (1962). *Arzneimitt.-Forsch.*, **12**, 431-435.
Kraus, L. (1957). *Pharmazie*, **12**, 693-695.
Lister, R. E. and Pride, R. R. A. (1959). *J. Pharm. Pharmacol.*, **11**, 278T-282T.
Mary, N. Y., Christensen, B. V. and Beal, J. L. (1956). *J. Amer. pharm. Ass. Sci. Ed.*, **45**, 229-232.
Paris, R. and Durant, M. (1956). *Ann. pharm. franç.*, **14**, 755-761.
Seel, E. (1919). *Arch. Pharm., Berl.*, **257**, 212-259.

SOME ASPECTS OF THE PHARMACOLOGY OF ORPHENADRINE

BY G. ONUAGULUCHI* AND J. J. LEWIS

From the Department of Materia Medica and Therapeutics and Division of Experimental Pharmacology, Institute of Physiology, University of Glasgow

Received December 14, 1962

Orphenadrine possesses muscle-relaxant activity resembling that of tubocurarine in preparations of the hen, rat and frog. It antagonises suxamethonium-induced contracture in the hen but increases the duration of depression of the twitch height. The bearing of these findings upon the anti-Parkinson's activity and other uses of orphenadrine is discussed.

CLINICAL experience has shown that the skeletal musculature is involved to a considerable degree in the rigidity and weakness of Parkinsonism. Despite this, relatively little attention has been paid to the effects of anti-Parkinson drugs on skeletal muscle. The most comprehensive study of the pharmacology of orphenadrine is that of Bijlsma and others (1956), but the effect upon skeletal muscle was not studied. The present investigation therefore considers some of the effects of orphenadrine on skeletal muscle with some observations upon its anticonvulsant action and effects upon spinal reflexes.

METHODS AND MATERIALS

Hen Gastrocnemius Muscle-Sciatic Nerve Preparation

The method used was based on that of Pelikan and his associates (1954). Hens weighing from 1.5 to 2.0 kg. were anaesthetised by 40 to 50 mg./kg. of sodium pentobarbitone given intravenously. The sciatic nerve on one side was exposed, shielded platinum electrodes placed on it and supra-maximal stimulation applied from a square wave generator at a frequency of 8/min., 25 to 50 V, pulse width 4 msec. In each experiment, voltage, pulse width and frequency were constant. The contractions were recorded using a Sherrington myograph lever. The sciatic nerve was crushed centrally to the electrodes.

Effect of orphenadrine on suxamethonium-induced contracture and neuromuscular block. Control responses were obtained with 12.5 to 25.0 $\mu\text{g./kg.}$ of suxamethonium. When these became reproducible, the effect upon subsequent responses of 1, 3, 5 or 6 mg./kg. of orphenadrine was investigated.

Effect of orphenadrine on tubocurarine-induced neuromuscular block. Control responses were obtained using 200 to 300 $\mu\text{g./kg.}$ of tubocurarine. It was usually necessary to give three similar doses before the degree of neuromuscular block was constant. After two similar control responses to the same dose of tubocurarine were obtained, 3 mg./kg. of orphenadrine were given and after 5 to 10 min., tubocurarine (200 to 300 $\mu\text{g./kg.}$) was repeated. The dose of tubocurarine was constant in each experiment.

* Present address: Department of Pharmacology, University College, Ibadan.

Effect of orphenadrine on the response to indirect tetanisation. The muscle was tetanised at 10 to 15 min. intervals by giving 30 sec. bursts of square impulses at a frequency of 1,400 to 1,500/min. After taking two or three control records, 3 mg./kg. orphenadrine was injected. The frequency of the tetanising current and the rest period between successive tetanisations were constant in each experiment.

Cat Gastrocnemius Muscle-Sciatic Nerve Preparation

The preparation was set up by conventional methods using 2 to 3.5 kg. cats, anaesthetised by 60 mg./kg. sodium pentobarbitone given intraperitoneally. Stimulation of the sciatic nerve was by supramaximal square impulses at 25 to 60 V, pulse width 3 to 4 msec. and frequency of 8/min. Tetanus was induced by increasing the frequency to 1,600/min. In each experiment voltage, frequency and pulse width were constant.

Effect of orphenadrine on suxamethonium-induced neuromuscular block. Suxamethonium 50–150 μ g./kg. was used, but the amount was constant for each experiment. At least two similar consecutive control responses were obtained before orphenadrine 3–6 mg./kg. was injected intravenously. 1–2 min. later a further dose of suxamethonium was administered and this was repeated when the contractions had returned to normal.

Effect of orphenadrine on tubocurarine-induced neuromuscular block. Control responses were obtained using tubocurarine 100 to 150 μ g./kg., repeating administration until the responses were constant. Orphenadrine 3 to 5 mg./kg. was then injected. After 2 min. tubocurarine was again given and administration repeated at intervals until control levels were attained.

Effects of orphenadrine on the response to indirect tetanisation. The muscle was tetanised at 15 min. intervals for 1 hr. using 30 sec. bursts of square impulses at 1,600/sec. After two control records had been obtained, orphenadrine 3 to 5 mg./kg. was given and tetanisation continued.

The Frog Rectus Abdominis Muscle Preparation

The muscle was set up in a 10 ml. bath containing frog Ringer's solution. Uniform, submaximal responses to acetylcholine were obtained using a 4 min. time cycle. Orphenadrine 15 to 25 μ g./ml. was given 15 sec. before the subsequent dose of acetylcholine.

The Isolated Rat Phrenic Nerve-diaphragm Preparation

The method was essentially that of Bülbring (1946) using a 100 ml. organ bath containing double glucose Tyrode's fluid at $29 \pm 0.5^\circ$ and Bell's (1952) electrode so that the muscle could be stimulated both directly and indirectly. Nerve stimulation was by square impulses; frequency 6 to 8/min. at 10 to 15 V and pulse width 0.5 to 1.0 msec. Direct stimulation of the muscle was at the same frequency but 30 to 50 V and pulse width 1.5 to 2.0 msec. In any experiment frequency, voltage and pulse width were constant. Drugs in aqueous solution were added to the bath and allowed to act for 3 min.

PHARMACOLOGY OF ORPHENADRINE

Effects of Orphenadrine on the Patellar and Crossed Extensor Reflexes in the Cat

Cats weighing between 2.0 and 5.5 kg. were anaesthetised by 50 mg./kg. sodium pentobarbitone given intraperitoneally, the hind brain and upper cervical cord destroyed, and maintained upon artificial respiration. The patellar reflex was elicited by stimulating the patellar tendon electrically at a frequency of 6/sec., 25 to 60 V and pulse width of 1.5 to 3.5 msec. The reflex responses were elicited for periods of from 2 to 3 min. with rest periods of 3 min. In any given experiment the period of stimulation, frequency, voltage and pulse width were constant. The crossed extensor reflex was elicited by stimulating supramaximally the right sciatic nerve and recording the contractions of the left quadriceps muscle. Single shocks from a square wave generator were employed at a frequency of 6 to 10/min., 10 to 40 V, pulse width 1.5 to 3.5 msec.

In any experiment frequency, voltage and pulse width were constant.

Effect of Orphenadrine on Leptazol and Electroshock Convulsions in Mice

Leptazol seizures were induced in female albino mice weighing between 18 and 26 g. The mice were divided into groups of 10. Four control groups received leptazol alone, each group receiving either 20, 30, 40 or 60 mg./kg. Other groups were pretreated with 1, 3 or 15 mg./kg. of orphenadrine, and leptazol at the above dose levels given 25 to 35 min. later. All drugs were given by injection into the tail vein. Electroshock seizures were induced by the method used by Ahmad and Lewis (1960) using the ear-clip electrodes of Hoyt and Rosvold (1951). Four groups of 20 female albino mice weighing between 15 and 21 g. were used. The current intensity was 17 mA in the groups treated with 3 or 12 mg./kg. of orphenadrine. 20 mA was employed in the groups which received 6 or 15 mg./kg. of orphenadrine. The current was not allowed to act for more than 5 sec. and was interrupted earlier if it produced tonic extension of the hind limbs. This was taken as the end-point. Seizure threshold was determined in each of the four groups of mice. Twenty-four hr. later the mice were given the appropriate dose of orphenadrine by injection into the tail vein and the electrical current applied 25 to 35 min. later.

RESULTS

In 3 out of 8 suxamethonium-treated hens, 3 mg./kg. of orphenadrine diminished the degree and duration of the contracture and increased the duration of the inhibition of the twitch height. In one hen this dose of orphenadrine itself produced a fall in the twitch height which lasted for about 20 min. In 6 hens, pretreatment with orphenadrine, 3 to 6 mg./kg., increased the degree and duration of the suxamethonium-induced block and in two preparations, 60 and 90 μ g./kg. of neostigmine antagonised a suxamethonium block after initial treatment with orphenadrine (Fig. 1). Pretreatment with 3 mg./kg. of orphenadrine increased the degree of muscle relaxation caused by tubocurarine. As a rule, both the intensity and duration of effect were increased but in some instances duration only

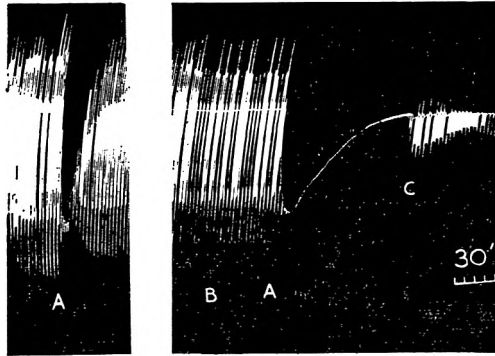


FIG. 1. Hen gastrocnemius muscle-sciatic nerve preparation. Pentobarbitone anaesthesia. Indirect stimulation by the sciatic nerve. Contractions downwards. Drugs administered intravenously.

At A, Suxamethonium, 12.5 μ g./kg. B, Orphenadrine, 3.0 mg./kg. C, Neostigmine, 90 μ g./kg.

was increased. Orphenadrine, 3 mg./kg., given alone did not alter the response to an indirect tetanisation.

In the cat, orphenadrine, 3 to 6 mg./kg., produced no muscular relaxation but antagonised the neuromuscular block induced by suxamethonium (Fig. 2). It increased the duration and amplitude of a tubocurarine block (Fig. 3) but did not alter the response to indirect tetarisation.

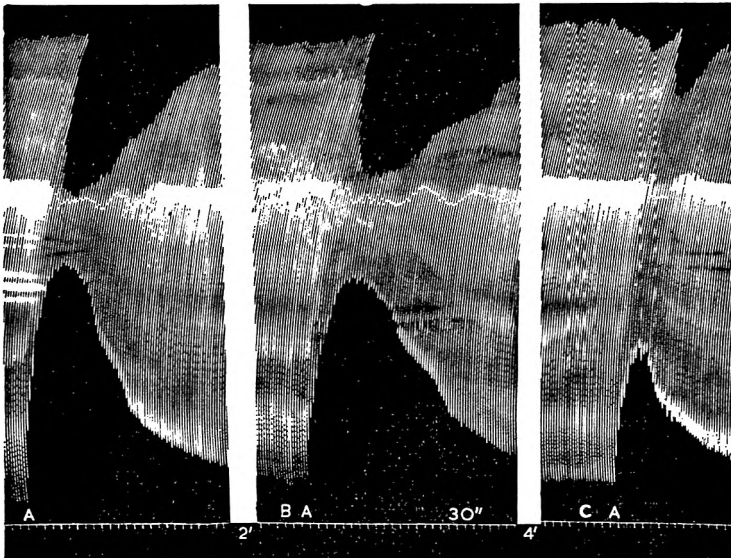


FIG. 2. Cat gastrocnemius muscle-sciatic nerve preparation. Pentobarbitone anaesthesia. Indirect stimulation by the sciatic nerve. Contractions downwards. Drugs administered intravenously.

At A, Suxamethonium, 150 μ g./kg. B, Orphenadrine, 3 mg./kg. C, Orphenadrine, 6 mg./kg.

PHARMACOLOGY OF ORPHENADRINE

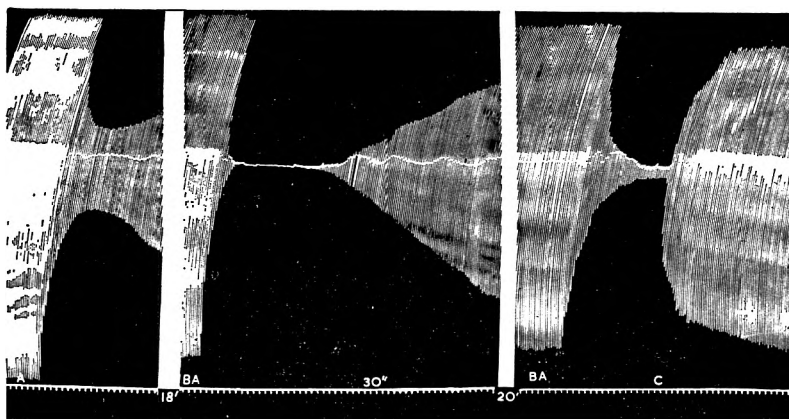


FIG. 3. Cat gastrocnemius muscle-sciatic nerve preparation. Pentobarbitone anaesthesia. Indirect stimulation by the sciatic nerve. Contractions downwards. Drugs administered intravenously.

At A, Tubocurarine, 125 $\mu\text{g./kg.}$ B, Orphenadrine, 3.0 mg./kg. C, Neostigmine, 100 $\mu\text{g./kg.}$

Orphenadrine, 25 $\mu\text{g./ml.}$, antagonised acetylcholine-induced contractions of the isolated frog rectus abdominis muscle. Recovery was slow despite frequent washing (Fig. 4). The effect of tubocurarine appeared to be additive with that of orphenadrine.

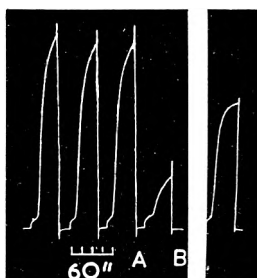


FIG. 4. Isolated frog rectus abdominis muscle. All unlabelled contractions due to 1 $\mu\text{g./ml.}$ of acetylcholine acting for 90 sec. Contraction A was preceded 1 min. earlier by orphenadrine, 2.5 $\mu\text{g./ml.}$

B, Time interval of 52 min. during which tissue was stimulated by acetylcholine 1 $\mu\text{g./ml.}$ at intervals of 4 min.

In the rat diaphragm, orphenadrine, 10 to 60 $\mu\text{g./ml.}$, reduced the twitch height following indirect stimulation. Orphenadrine was from one-eighth to one-fifteenth as potent as tubocurarine on this preparation. Following complete paralysis with orphenadrine the muscle still responded to direct stimulation. Recovery was prolonged and usually incomplete except at very low dose levels.

In spinal cats the effects of orphenadrine on the patellar tendon and crossed extensor reflexes were variable. In some instances the magnitude of the response was slightly reduced; in others it was slightly increased;

in some there was no change. In some animals, muscular spasms which appeared within 1 min. of the injection were noted. These were usually short-lived but were sometimes associated with an increase in the magnitude of the reflex response (Fig. 5).

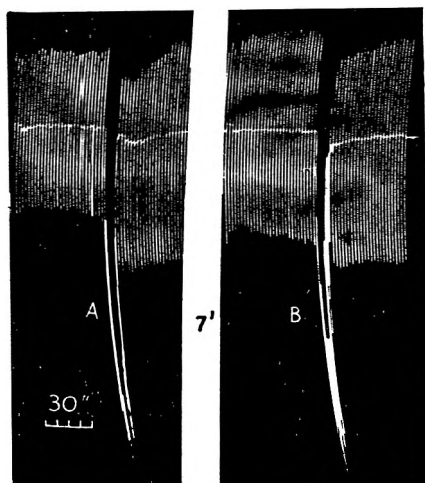


FIG. 5. Spinal Cat. Effect of orphenadrine on the crossed extensor reflex. At A, Orphenadrine, 3 mg./kg. B, Orphenadrine, 5 mg./kg.

At dose levels of 1, 3 and 15 mg./kg., orphenadrine did not alter the number of animals convulsing after dosage with leptazol although the intensity of the tonic phase was reduced. At the highest dose level there was a reduction in mortality. 3 and 6 mg./kg. of orphenadrine had no effect on the incidence of electroshock seizures in mice but reduced this at 12 and 15 mg./kg.

DISCUSSION

The experiments on the frog rectus abdominis muscle and the rat diaphragm indicate that orphenadrine has some muscle relaxant activity.

The antagonism to suxamethonium-induced block and potentiation of tubocurarine block in the cat also demonstrated that despite its very low potency some changes may take place at the neuromuscular synapse when this drug is given. It is possible that orphenadrine acts at the synapse by competing with acetylcholine and other depolarising drugs used. It is also possible that these effects are due in some part to a direct action on the muscle or, but more unlikely, to interneuronal block. As judged from these experiments the effects of orphenadrine last for about an hour.

In the hen, the actions of orphenadrine are difficult to assess. Orphenadrine-prolongation of suxamethonium block is an action not unlike that reported for tubocurarine by Crema, Scognamiglio and Bovet (1959). In the hen, orphenadrine appears therefore to mimic the actions of a non-depolarising neuromuscular blocking agent, although there is no direct evidence that it has in fact a tubocurarine-like action. In the rat

PHARMACOLOGY OF ORPHENADRINE

diaphragm and frog rectus abdominis muscle preparations, the actions of orphenadrine are more prolonged than those of tubocurarine but on these preparations it is much less potent.

In mice, even rather large doses of orphenadrine (15 mg./kg.) had no apparent effect upon the convulsant activity of leptazol. This is in contrast to the results of Cronheim (1958) who found that orphenadrine lowered the threshold for leptazol-induced convulsions. On the other hand, it was noted that in the first minute after 15 mg./kg. of orphenadrine, the mice were more restless, and in two instances orphenadrine-treated mice died from convulsions within 2 min. At this dose level orphenadrine may therefore have an initial excitatory or facilitatory action which soon changes into one of depression. This high dose of orphenadrine did however reduce the mortality to leptazol and the intensity of the tonic phase of the seizure and the effect of electroshock. At this level orphenadrine may inhibit seizure spread in the cerebrospinal axis but peripheral muscle-relaxant activity may also contribute to the anticonvulsant activity.

In Parkinson's disease the facilitatory system of the reticular formation is over-active and this is associated with a great increase in the gamma efferent discharge to the muscle spindles. This, through the servo-mechanism of Kuffler, Hunt and Quilliam (1951) leads to hypertonia of the skeletal muscles. The reticular formation can influence the spinal cord and, through the spinal internuncial cells, the skeletal muscles. Rigidity or hypertonia can thus be modified or abolished by depression of the activity of the reticular formation, basal ganglia, diencephalon or cerebellum, of the spinal interneurons or, by breaking the reflex arc of the servo-mechanism responsible for the hypertonia.

Orphenadrine reduces rigidity and hypertonia in patients suffering from Parkinson's disease. Suxamethonium in subparalytic doses increases the efferent discharge from the muscle spindles (Granit, Skoglund and Thesleff, 1953). Smith and Eldred (1961) have suggested that this effect is due to depolarisation of the end-plate region of the intrafusal fibres. It is possible, therefore, that the beneficial action of orphenadrine in Parkinson's disease is in part due to a depressant effect on muscle spindle discharge. Local anaesthetic activity (Bijlsma and others, 1956) may also play a part in the anti-Parkinson's activity of orphenadrine. The euphoriant action (Onuaguluchi, 1961), vasodilation of skeletal muscle (Bijlsma and others, 1956) and an effect upon phosphate metabolism (Van Petten and Lewis, 1962) may also be involved. The experiments reported in this paper have led to trial of orphenadrine to prevent muscular pain following suxamethonium.

Acknowledgments. One of us (G.O.) is much indebted to Professor S. Alstead, Regius Professor of Materia Medica and Therapeutics in the University of Glasgow, for giving him the opportunity to take part in this study. We are grateful to Miss Gladys Marren for technical assistance and to Dr. G. Schmeidler of the Camden Chemical Company for a supply of orphenadrine. Our thanks are due to Dr. T. C. Muir for helpful discussions.

G. ONUAGULUCHI AND J. J. LEWIS

REFERENCES

- Ahmad, K. and Lewis, J. J. (1960). *J. Pharm. Pharmacol.*, **12**, 163-174.
- Bell, G. H. (1952). *Experimental Physiology*, 5th Edition, pp. 36-44. John Smith & Son (Glasgow) Ltd.
- Bijlsma, U. G., Harms, A. F., Funcke, A. B. H., Tersteeg, H. M. and Nauta, W. Th. (1956). *Arch. int. Pharmacodyn.*, **106**, 332-369.
- Bülbring, E. (1946). *Brit. J. Pharmacol.*, **1**, 38-61.
- Crema, A., Scognamiglio, W. and Bovet, D. (1959). *Arch. int. Pharmacodyn.*, **122**, 152-167.
- Cronheim, G. (1958). *J. Pharmacol.*, **122**, 16A.
- Granit, R., Skoglund, S. and Thesleff, S. (1953). *Acta physiol. scand.*, **28**, 134-151.
- Hoyt, R. and Rosvold, H. E. (1951). *Proc. Soc. exp. Biol.*, N.Y., **78**, 582-583.
- Kuffler, S. W., Hunt, C. G. and Quilliam, J. P. (1951). *J. Neurophysiol.*, **14**, 29-54.
- Onuaguluchi, G. (1961). Ph.D. Thesis, Glasgow University.
- Pelikan, E. W., Smith, C. M. and Unna, K. R. (1954). *J. Pharmacol.*, **111**, 30-42.
- Smith, G. M. and Eldred, E. (1961). *Ibid.*, **131**, 237-242.
- Van Petten, G. R. and Lewis, J. J. (1962). *Ibid.*, **136**, 372-377.

A NOTE ON THE PREPARATION OF RICINOLEIC ACID BY UREA COMPLEXING

BY DIPTISH CHAKRAVARTY AND ARUN BOSE

From the Research Division, Smith, Stanistreet & Co. Ltd., Calcutta-14

Received August 14, 1962

Some fatty acids were separated from hydrolysed castor oil by successive additions of urea. The method was utilised to prepare ricinoleic acid of B.P. 1948 standard from castor oil.

DURING recent work we required ricinoleic acid conforming to the B.P. 1948 standard. One of the simplest methods of obtaining the acid is by the hydrolysis of castor oil, but the product so obtained does not conform to the B.P. 1948 standard. The distillation of crude ricinoleic acid under reduced pressure involves the risk of polymerisation, the method being commercially uneconomical. Another method of purification utilises the process of solidification which is very slow because of the oily nature of the material, and a further disadvantage is that the temperature has to be maintained below 4° throughout the operation. Moreover, the congealing point of crude ricinoleic acid is close to that of the more pure form.

Mehta and Dabhade (1959) applied the urea complexing method to the separation of different constituents of chaulmoogra oil, which was otherwise difficult. The various fatty acids were regenerated from urea complexes by hydrolysis with hydrochloric acid. The method (Sinha, Chakrabarty and Chakrabarty, 1957) utilises the fact that in urea-adduct formation, the influence of melting-point and solubility are subordinate to the influence of unsaturation. With increasing saturation in the conjugated acids there is a marked tendency towards adduct formation with lesser concentrations of urea. Increase in the unsaturation and chain length interferes with the complex formation. Based on the above findings we have been able to obtain ricinoleic acid of B.P. 1948 quality from castor oil.

EXPERIMENTAL

Hydrolysis of castor oil. Castor oil (50 g.) was added to alcoholic caustic potash solution (25 g. in 225 ml. ethanol) and the solution was refluxed for 16 hr. or until a test portion of the reaction mixture was completely soluble in water. The ethanol was distilled from the reaction mixture and the residue dissolved in water (200 ml.), washed with ether, decolourised with charcoal and filtered. The filtrate was acidified with hydrochloric acid and the separated oil taken up in ether (200 ml.). The ethereal solution was washed with water and dried over anhydrous sodium sulphate. On removal of ether, a light-brown liquid (34 g.) was obtained.

Urea complexing. Hydrolysed castor oil (25 g.) thus obtained was dissolved in anhydrous ethanol (250 ml.). Urea (30 g.) was dissolved in the above solution by refluxing over a water-bath. The solution on cooling

DIPTISH CHAKRAVARTY AND ARUN BOSE

yielded a colourless crystalline solid, which was filtered off. The solid was one fraction of the fatty acids. After diluting the filtrate to 250 ml. with anhydrous ethanol, more urea (25 g.) was added and dissolved by refluxing, and the solution cooled to yield a colourless crystalline solid, which was filtered off and dried. This solid was suspended in water (150 ml.) acidified with hydrochloric acid and heated on a water-bath for 15 min., the mixture was cooled, the separated oil taken up in ether, and the ether solution washed with water and dried over anhydrous sodium sulphate. On removal of ether, a yellowish-white liquid (5 g.), which is the ricinoleic acid, was obtained. The experiment was repeated with different samples of hydrolysed castor oil containing crude ricinoleic acid. The analytical data on three samples of ricinoleic acid obtained by this method are recorded in Table I.

TABLE I

	B.P. 1948 specification	Sample No. 1	Sample No. 2	Sample No. 3
Description ..	Yellow or yellowish-brown viscous liquid	Yellowish-white liquid	Yellowish-white liquid	Yellowish-white liquid
Solubility	Insoluble in water; soluble in ethanol	Conforms	Conforms	Conforms
Acid value	Not less than 175	184.9	184	182.6
Freezing point ..	Does not congeal to a solid mass until cooled below 4°	Congeals at about 2°	Congeals at about 2°	Congeals at about 2°
Iodine value	85 to 91	89.6	87.7	88.1
Refractive index at 40°	1.462 to 1.468	1.464	1.462	1.467

From these results, it is evident that the method can be applied to obtain ricinoleic acid of B.P. 1948 standard.

Acknowledgement. Our thanks are due to Mr. B. Mitra for the analysis of samples.

REFERENCES

- Mehta, T. N. and Dabhade, S. B. (1959). *Grasas Yaceites (Seville, Spain)*, **10**, 24-26.
 Sinha, S., Chakrabarty, S. R. and Chakrabarty, M. M. (1957). *J. Ind. Chem. Soc. (Ind. & News Ed.)*, **20**, 104-105.

A NOTE ON THE STABILITY OF THE TRIFLUOROMETHYL GROUP OF BENDROFLUMETHIAZIDE IN RATS

BY G. HASSELMANN AND K. ROHOLT

From the Biological Department, Leo Pharmaceutical Products, Copenhagen, Denmark

Received November 2, 1962

The effect of bendroflumethiazide on the teeth of rats on a carious diet was examined and compared to that of sodium fluoride to determine whether release of fluoride takes place from the trifluoromethyl group of the substance. No detectable fluoride was released.

SINCE 1958 a series of fluorine-containing diuretics have appeared, the fluorine being incorporated in the molecules as a trifluoromethyl group ($-CF_3$). Chemical investigations have shown that the fluorine atoms in this group are very firmly bound to the carbon atom (Kobinger, Lund and Roholt, 1960). The stability of the trifluoromethyl group in bendroflumethiazide was examined *in vivo* since release of fluoride could lead to a chronic fluorine poisoning in long term treatment with this diuretic.

Chronic fluorosis causes striking effects on the teeth which show alterations in the calcification of the enamel known as mottling, while administration of fluoride in amounts too small to produce mottling renders the teeth more resistant to caries. These observations have been used to detect the possible release of minute amounts of fluoride from the drug.

MATERIALS AND METHODS

The method of Pindborg (1958) was used. Albino rats of the Leo Strain received *ad libitum* from the age of two months the highly cariogenic diet used by Stephan and others (1952). As an extra supplement the animals had fresh fruit and B-vitamins once weekly. They were divided into 4 groups each consisting of 10 males and 10 females.

Group I served as control.

Group II received orally by stomach tube 15 mg. bendroflumethiazide suspended in 0.2 ml. of sucrose solution per rat per day (except Sundays). This dose has a calculated fluorine content of 2 mg.

Group III was given 2 mg. of sodium fluoride in 0.2 ml. of sucrose solution by stomach tube per animal daily (except Sundays).

Group IV had 22 mg. of sodium fluoride added to each litre of drinking water, which corresponds to an average daily intake of approximately 0.9 mg. of sodium fluoride.

After 85 days of administration the rats were killed. The heads were placed in dilute NaOH solution and all soft tissue was removed. The molars were examined microscopically using bright illumination and a sharp dental explorer and the degree of caries was scored following the principles of Gustafson and others (1952). Each carious tooth was given a severity score of 1, 2 or 3 indicating a caries attack of superficial, medium or extensive type. Each rat was scored individually and the average score for each group determined.

RESULTS

In the control group, the diet produced caries in all but two animals. The number of carious teeth per rat was 2.1, the average caries severity score 2.6.

In group II, receiving bendroflumethiazide, all animals but one developed caries. The average number of carious teeth was 3.1 and the average severity score 4.1.

A marked reduction in caries was found in groups III and IV, the average number of carious teeth being 0.4 and 1.0 and the average caries severity score being 0.5 and 1.1 respectively.

Within the same group no significant difference was seen between males and females neither with regard to number of carious teeth nor to caries severity score.

It was noted that 93 per cent of the carious teeth were found among the molars of the mandible.

In group III the enamel of all incisors had a banded appearance, each white band corresponding to each dose of sodium fluoride. In marked cases we were able to locate the region of growth on Sundays (no administration) by means of the broader normal orange pigmented bands. No such rings were seen in group II.

CONCLUSION

The results clearly demonstrate the caries inhibiting effect of fluoride on rat molars (Groups III and IV). Since no inhibition was observed in rats treated with bendroflumethiazide (group II) it can be concluded that no fluoride detectable by this method was released from the trifluoromethyl group of the drug.

REFERENCES

- Gustafson, G., Stelling, E. and Brunius, E. (1952). *Odont. Tidskr.*, **60**, 101-154.
Kobinger, W., Lund, F. and Roholt, K. (1960). *J. Irish med. Ass.*, **47**, 81-84.
Pindborg, J. J. (1958). *Acta odont. Scandinav.*, **16**, 383-388.
Stephan, R. M., Fitzgerald, R. J., McClure, F. J., Harris, M. R. and Jordan, H. (1952). *J. dental. Res.*, **31**, 421-427.

A NOTE ON THE SYNTHESIS OF ESTERS OF N-METHYLPYRROLIDINYALKANOLS

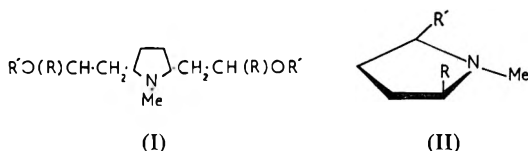
BY F. PERKS AND P. J. RUSSELL

*From the Pharmaceutical Chemistry Laboratory, Department of Pharmacy and
Physiology, College of Technology, Portsmouth*

Received October 31, 1962

The preparation of a series of esters of 1-methyl-2,5-di-2-hydroxy-2-phenylethylpyrrolidine is described. They are assigned *trans* configurations on the basis of their pK_a values.

ESTERS of *N*-methylpyrrolidinylalkanols of the type I (where $R = Me$ and $R' = Ph \cdot CO-$, $Ph \cdot CH : CH \cdot CO-$, $p-NH_2 \cdot C_6H_4 \cdot CO-$, $p-MeO \cdot C_6H_4 \cdot CO-$ and $Ph \cdot NH \cdot CO-$) have been described by Linnell and Perks (1960 a, b), and found to possess local anaesthetic activity.



It was therefore of interest to prepare a similar series of compounds but having $R = Ph-$, in order to determine the effect of replacing the methyl group of the alkanol side chains by the phenyl group.

1-Methyl-2,5-diphenacylpyrrolidine hydrochloride was prepared by the method of Schöpf and Lehmann (1935) and, after conversion to the free base, reduced with lithium aluminium hydride to 1-methyl-2,5-di(2-hydroxy-2-phenylethyl)-pyrrolidine. This was treated in acetone solution with the appropriate acid chloride and excess sodium hydroxide solution to give the pyrrolidinylalkanol esters (I; $R = Ph-$; $R' = Ph \cdot CO-$, $Ph \cdot CH : CH \cdot CO-$, $p-MeO \cdot C_6H_4 \cdot CO-$ and $p-NO_2 \cdot C_6H_4 \cdot CO-$). These were all viscous oils and were converted to their picrates for analysis. No soluble salts suitable for pharmacological testing could be prepared.

EXPERIMENTAL

All m.ps. are uncorrected. Microanalyses are by Mr. G. S. Crouch, School of Pharmacy, London.

1-Methyl-2,5-diphenacylpyrrolidine hydrochloride, prepared according to Schöpf and Lehmann (1935) had m.p. 200° and this was converted into the free base, m.p. 62° , by treatment with potassium hydroxide solution.

1-Methyl-2,5-di(2-hydroxy-2-phenylethyl)pyrrolidine. A suspension of 1-methyl-2,5-diphenacylpyrrolidine (75 g.) in dry ether (1.5 litre) (in which it is only slightly soluble), was stirred vigorously under reflux whilst a slurry of lithium aluminium hydride (10 g.) in dry ether (250 ml.) was added in small portions. The mixture was then refluxed and stirred

for 4 hr. and allowed to stand overnight. After destroying the excess lithium aluminium hydride with ethyl acetate, there was isolated in the usual manner 72 g. of the desired product as a yellow, viscous oil, b.p. 202–205° at 0.25 mm. Hg.

TABLE I
PICRATES OF PYRROLIDINYL ALKANOL ESTERS

Ester	m.p. °C of picrate	Analysis of picrate					
		Found (per cent)			Required (per cent)		
		C	H	N	C	H	N
Dibenzoate	185° (decomp.)	64.4	5.2	7.3	64.6	5.0	7.35
Dicinnamate	213° (decomp.)	66.2	5.1	6.8	66.3	5.2	6.9
Di- <i>p</i> -methoxybenzoate	201–3° (decomp.)	62.5	5.4	7.0	62.7	5.2	6.8
Di- <i>p</i> -nitrobenzoate	233–4° (decomp.)	57.4	4.4	9.9	57.7	4.3	9.85

The following *pyrrolidinylalkanol esters* (see Table I) were prepared from the above pyrrolidinylalkanol and the appropriate acid chloride (20 per cent excess) in acetone solution and an excess of sodium hydroxide solution. The basic esters were isolated by ether extraction, drying (Na_2SO_4), and removal of the ether. In all instances the products were viscous oils which were dissolved in dry benzene and passed through a short column of alumina, followed by removal of the solvent. For analyses the picrates were prepared from alcoholic solutions of the ester and picric acid. The di-*p*-nitrobenzoate could not be reduced to the corresponding aminobenzoate, all attempts giving intractable tars.

TABLE II
 pK_a VALUES OF PYRROLIDINE DERIVATIVES (II) $\text{R} = \text{R}'$

Series A		Series B		Series C	
No.	pK_a	No.	pK_a	No.	pK_a
1	CO_2Et 4.3	4	CH_2COMe 8.0	7	$\text{CH}_2\text{CO}\cdot\text{Ph}$ 6.9
2	CH_2OH 8.5	5	$\text{CH}_2\text{CHMe}\cdot\text{OH}$ 9.3	8	$\text{CH}_2\text{CHPh}\cdot\text{OH}$ 7.8
3	CH_2OBz 5.2	6	$\text{CH}_2\text{CHMe}\cdot\text{OBz}$ 7.0	9	$\text{CH}_2\text{CHPh}\cdot\text{OBz}$ 6.8

Measurement of pK_a values. pK_a values were measured using a Cambridge pH meter and glass electrode with calomel reference electrode. Solutions of the bases in 30 per cent ethanol were titrated with 0.1N hydrochloric acid. Graphs were plotted of the pH of the solution against volume of titrant added and the pK_a values calculated from the points of half-neutralisation.

DISCUSSION

The present series of compounds is believed to have a *trans* configuration like the previous series (Linnell and Perks (1960 a, b), on the basis of their pK_a values (see Table II). This was to be expected, since the parent compound ($\text{II.R} = \text{R}' = -\text{CH}_2\cdot\text{CO}\cdot\text{Ph}$) is prepared by a similar method to the corresponding diketone of the previous series ($\text{II, R} = \text{R}' = -\text{CH}_2\cdot\text{CO}\cdot\text{Me}$).

ESTERS OF *N*-METHYLPYRROLIDINYLLALKANOLS

The compounds of series A, included for comparison, have been shown to have the *cis* configuration and those of series B to have the *trans* configuration by considering proton addition to the ring nitrogen in terms of steric factors and hydrogen bonding (Linnell and Perks, 1960a).

Of the present series C, compound No. 7 is a comparatively strong base (pK_a 6.9) though somewhat weaker than compound No. 4 (pK_a 8.0) due to the more effective shielding of the basic centre by the bulkier substituent. This may be compared with the much weaker base No. 1 (pK_a 4.3) where there is much more efficient shielding by two much smaller substituents in the *cis* configuration. Compound No. 8 (pK_a 7.8) is a stronger base than No. 7, because of the base-strengthening hydrogen bonding effect, the steric factor being very similar in each case. In compound No. 9, the hydrogen bonding effect is lost and there is a fall in base strength (pK_a 6.8). This is very near to the value for compound No. 6 (pK_a 7.0) and it would seem that with substituting groups of such size and complexity, the steric effect has reached a maximum, larger groups being no more effective in shielding the basic centre.

Compounds of series C therefore, 1-methyl-2,5-diphenacylpyrrolidine and its derivatives are assigned the *trans* configuration.

REFERENCES

- Linnell, W. H. and Perks, F. (1960a). *J. chem. Soc.*, **213**, 1036-1040.
Linnell, W. H. and Perks, F. (1960b). *J. Pharm. Pharmacol.*, **12**, 95-98.
Schöpf, C. and Lehmann, G. (1935). *Annalen*, **518**, 1.

LETTERS TO THE EDITOR

Mouse Strain Difference in Response to Antihistamine Drugs

SIR,—It may appear as a truism to state that the species of animal sensitive to histamine challenge respond more readily to the protective action of antihistamines but differences in the responses of mouse strains serve to illustrate that such diversity may be of genetic origin.

Mice from five strains (one random bred, LAC greys, two inbred, C57BR/cd and CBA, and their reciprocal crosses) were sensitised to histamine with pertussis vaccine. The controls were challenged directly with histamine acid phosphate while the test mice first received a subcutaneous dose of one of three antihistamine compounds and were subsequently challenged. Adult mice between six and eight weeks old were used, and where available were distributed in equal numbers of the sexes and equal numbers of control and test mice. The differences between the numbers of mice surviving in the test groups and in the control groups were calculated as mice surviving due to the action of the antihistamine drug.

Results in Table I are in the order of strain sensitivity to histamine that had been previously determined (Brown 1959) and the challenge dose of histamine used for each strain was at least 2.5 times the approximate average lethal dose. The absolute histamine sensitivity for each strain varies with the sensitisation response to histamine produced by the batch of pertussis vaccine used. The hybrid strains and the LAC greys have an average lethal dose to histamine of about 100 mg./kg. body weight. To a larger dose of histamine, 250 mg./kg. mouse, the LAC greys are slightly less sensitive than the hybrid strains and the effect of antihistamine, particularly that of the larger doses, upon the responses, is perhaps not so marked and thus in accordance with the known effects in other species.

TABLE I

THE EFFECT OF ANTIHISTAMINE DRUGS GIVEN SUBCUTANEOUSLY TO MICE 20 MIN. BEFORE CHALLENGE WITH HISTAMINE ACID PHOSPHATE

Strain	Histamine acid phosphate mg./kg. mouse	No. of mice per group	Surviving control mice		Mice surviving as a result of antihistamine dose in mg. per mouse											
					Tripeleannamine				Chlorpheniramine				Diphenhydramine			
					0.08		0.40		0.02		0.10		0.08		0.40	
Nos.	per cent	Nos.	per cent	Nos.	per cent	Nos.	per cent	Nos.	per cent	Nos.	per cent					
C57BR/cd	100	21	1	5	11	52	19	91	1	5	18	86	8	38	20	95
BRCF ₁	250	38	4	11	16	42	33	87	19	50	33	87	24	63	34	90
CBRF ₁	250	41	5	12	23	56	33	81	17	42	33	81	21	51	35	85
LAC greys	250	37	10	27	18	49	28	76	15	41	26	70	17	46	28	76
CBA	800	44	21	48	5	11	3	7	5	11	15	34	2	5	6	14

A consideration of the inbred strains and their genetically similar reciprocal crosses, shows that the insensitive strain CBA is little protected against histamine by any antihistamine drug, while the other strains are in the main well protected. An exception is shown by the C57BR/cd mice against the lower dose of chlorpheniramine. A five-fold increase in the concentration of this drug results in a seventeenfold increase in response. Such a demonstration of precision suggests that chlorpheniramine action is specific against histamine, and that C57BR/cd mice would be exceptionally valuable for its assay.

LETTERS TO THE EDITOR

Several deductions may be made from the general responses to antihistamine of drugs of the inbred strains and their hybrids. First, the unresponsiveness of the CBA strain and the responsiveness of the hybrids and C57BR/cd strain indicate that this reaction is controlled by an incompletely dominant trait. Non-response, as it is confined to one parent and does not appear in the progeny, may be controlled by a recessive gene in the same way as non-anaphylactoid reaction (Harris and West, 1961). Second, the same pattern of response occurs in all species; those animals easily sensitised to histamine are well protected by antihistamine. Finally, the differentiation between antihistamines by the homozygous C57BR/cd strain may point to a relation between homozygosity and specificity, suggesting that at some point in this particular reaction a single gene may be involved.

Laboratory Animals Centre,
M.R.C. Laboratories,
Woodmansterne Road,
Carshalton, Surrey.
March 6, 1963

ANNIE M. BROWN.

REFERENCES

- Brown, A. M. (1959). *L.A.C. Collected Papers*, 8, 9-16.
Harris, J. M. and West, G. B. (1961). *Nature (Lond.)*, 191, 399-400.

Gamma Irradiation of *Bacillus subtilis* Spores

SIR,—When aqueous suspensions of *Bacillus subtilis* spores were exposed to gamma irradiation from a Cobalt-60 source and subsequently stored at 0° and 20° more survived at 20° than at 0°.

Also the slopes of the lines relating dose to numbers of survivors were not parallel, but converged at approximately total survival at no dose of radiation. Thus spores subsequently stored at 20° survived larger doses of gamma radiation than the spores subsequently stored at 0°.

These results are summarised in Table I.

TABLE I
DIFFERENCE IN SLOPES OF LOG PER CENT SURVIVOR/DOSE REGRESSIONS AFTER STORAGE FOR ONE MONTH AT 0-4° AND 20-26°

Temperature	Correlation coefficient	Regression coefficient (rad. 10 ⁻⁵)	'D' value (rad. 10°)
0-4°	-0.9841	-0.5425	1.83
20-26°	-0.9950	-0.4493	2.23

('D' value = decimal reduction factor)

Calculated 'd' (Bailey, 1959) = 2.284

Tabulated 't' (P = 0.05) = 2.228

There is therefore a significant difference in slope.

When an aqueous suspension of *B. subtilis* spores is heated at 60° for 3 min., or 60° for 15 min., or 90° for 5 min., there is a progressive increase in the number of spores which produce colonies on agar plates. But if a second sample of the spore suspension was first irradiated and then heated in the same way, there was a decrease in the number of spores producing colonies on agar plates (Table II.)

It is well-known that heating at sub-lethal temperatures causes dormant spores to germinate with a resultant increase in viable count (Desrosier and Heiligman, 1956; Curran and Evans, 1945 and 1947). This response varies

LETTERS TO THE EDITOR

with the temperature and duration of heating (Halvorson and Church, 1957; Murrell, 1961).

TABLE II
VARIATION IN RECOVERY OF *B. subtilis* AFTER POST-IRRADIATION HEAT-SHOCK

Dose (rad.)	Percentage increase over unheated recovery		
	3 min./60°	15 min./60°	5 min./90°
0	59	64	82
4 × 10 ²	37	27	-25
6 × 10 ²	13	17	-34

The results indicate that pre-irradiation sensitises *B. subtilis* spores to heating normally sub-lethal and used to activate dormant spore suspensions.

Department of Pharmaceutics,
School of Pharmacy,
University of London,
29/39, Brunswick Square,
London, W.C.1.
March 26, 1963

A. M. COOK
T. A. ROBERTS

REFERENCES

- Bailey, N. J. T. (1959). *Statistical Methods in Biology*. London: English Universities Press Ltd.
 Curran, H. R., and Evans, F. R. (1945). *J. Bact.*, **49**, 335-346.
 Curran, H. R., and Evans, F. R. (1947). *Ibid.*, **53**, 103-113.
 Desrosier, N. W., and Heiligman, F. (1956). *Food Res.*, **21**, 54-62.
 Halvorson, H., and Church, B. D. (1957). *Bact. Rev.*, **21**, 112-131.
 Murrell, W. G. (1961). *Microbial Reaction to Environment*, 11th Symposium Soc. gen. Microbiol. (ed. Meynell, G. G. and Gooder, H.), p. 100. Cambridge University Press.

The Antibacterial Action of Glycine

SIR,—It is known that some D-amino-acids exert an inhibitory effect on certain Gram-negative bacteria, and that in the presence of sucrose as a stabilising agent, morphological variants can be preserved (Jeynes, 1957; Welsch, 1958; Lark and Lark, 1959; 1961).

The term "spheroplasts" has been suggested for these variants which might retain at least a portion of the original cell wall (Brenner and others, 1958) and which have also been induced by treating *Escherichia coli* with various penicillins (Russell 1962; Turner and Russell 1962).

In the present preliminary report, an investigation has been made into the quantitative aspects of the effect of glycine on *E. coli* in a nutrient medium in the presence or absence of sucrose and Mg⁺⁺ ions. These ions have previously been found to be essential in stabilising penicillin-induced spheroplasts (Lederberg 1956; Hugo and Russell 1960).

In our experiments 0.1 ml. of an overnight 37° broth culture of the organism was added to 10 ml. tubes of nutrient broth containing 0.33 M sucrose and 0.25 per cent w/v MgSO₄·7H₂O, and varying concentrations of glycine. After incubation of all tubes at 37° for 4 hr., three samples were examined.

(1) 1 ml. was added to 9 ml. of sterile water to lyse any spheroplasts present. Further serial dilutions were made if necessary, 1 ml. samples being finally plated into 10 ml. of nutrient agar.

LETTERS TO THE EDITOR

(2) 1 ml. was added to 9 ml. of 0.33 M sucrose: Mg⁺⁺ broth with the aim of protecting any spheroplasts from osmotic shock, and after further serial dilution if necessary in this medium, 1 ml. samples were plated into 10 ml. of 0.33 M sucrose: Mg⁺⁺ agar.

(3) Aliquots were examined by phase-contrast microscopy. The results of a typical experiment are shown in Table I.

TABLE I
THE EFFECT OF GLYCINE ON *E. coli*

Glycine concentration mg./ml.	Count of survivors in agar per ml.	Count of survivors in 0.33 M sucrose Mg ⁺⁺ agar per ml.	Presence of spheroplasts
0	> 10 ⁷	> 10 ⁷	—
20	1.9 × 10 ⁴	4.3 × 10 ⁴	++
25	2.4 × 10 ⁴	1 × 10 ⁴	+
30	8.5 × 10 ³	2.1 × 10 ³	+

++ Optimum number of spheroplasts.
+ Few spheroplasts.
— No spheroplasts.

The optimum concentration of glycine to induce spheroplasts, was 20 mg./ml. At 15 mg./ml. of glycine, spheroplasts, bizarre forms and rods could be observed microscopically. Above 25 mg./ml. the number of spheroplasts dropped sharply, although the bacteria were still killed, as indicated by the survivor counts. In agar containing sucrose and Mg⁺⁺ ions, survivor counts were slightly higher than those obtained in ordinary nutrient agar alone. This suggests that some spheroplasts, at least, were able to survive in the presence of the stabilisers, and produce colonies.

In nutrient broth, to which sucrose and MgSO₄·7H₂O had not been added, no spheroplasts were induced by any concentration of glycine.

The Welsh School of Pharmacy,
Welsh College of Advanced Technology,
Cardiff.
March 26, 1963

R. V. JOHN
A. D. RUSSELL

REFERENCES

- Brenner, S., Dar^c, F. A., Gerhardt, P., Jaynes, M. H., Kandler, O., Kellenberger, E., Klieneberger-Nobel, E., McQuillen, K., Rubio-Huertos, M., Salton, M. R. J., Strange, R. E., Tomcsik, J. and Weibull, C. (1958). *Nature, Lond.*, **181**, 1713-4.
Hugo, W. B. and Russell, A. D. (1960). *J. Bacteriol.*, **80**, 436-440.
Jaynes, M. H. (1957). *Nature, Lond.*, **180**, 867.
Lark, C. and Lark, K. G. (1959). *Canad. J. Microbiol.*, **5**, 369-379.
Lark, C. and Lark, K. G. (1961). *Biochim. et Biophys. Acta*, **49**, 308-322.
Russell, A. D. (1962). *J. Pharm. Pharmacol.*, **14**, 390-392.
Turner, T. D. and Russell, A. D. (1962). *J. Pharm. Pharmacol.*, **14**, 395-396.
Welsch, M. (1958). *Schweiz Z. allgem. Pathol. u. Bakteriologie*, **21**, 741-768.

LETTERS TO THE EDITOR

Diffusion of Salts Through a Lipo-protein Interface

SIR,—The rates of diffusion of some salts through an interface formed between sols of lipid and protein have been measured every hour using a capillary cell and a conductivity apparatus. The interface was formed as previously described by Saunders (1963). The salt under examination was added to each separate sol in differing concentrations and the diffusion rate across the interface was followed by noting the changes of conductivity with time between electrodes set 5 mm. below the boundary.

Sodium chloride diffuses very slowly across the interface between a 10 per cent bovine serum albumin sol and a sol containing 10 per cent of lecithin and 5 per cent of cholesterol, dispersed ultrasonically. The rate is particularly slow in the presence of calcium chloride (Saunders, 1963).

TABLE I

EFFECT OF CHOLINE SALTS ON THE DIFFUSION OF SODIUM CHLORIDE ACROSS A LIPO-PROTEIN INTERFACE

Δ is the ratio of the change in conductivity between the electrodes to the initial difference in conductivity between the two sols. Temperature 24°.

Time hr.	Δ					
	A	B	C	D	E	F
1	0	0.03	0.01	0.02	0	0.13
2	0	0.04	0.10	0.03	0	0.19
4	0.01	0.09	0.16	0.12	0.02	0.27
8	0.02	0.12	0.21	0.20	0.04	0.32
12	0.03	0.13	0.22	0.24	0.06	0.34

- A. Lower sol., 10 per cent bovine serum albumin, 0.02N NaCl, 0.001N CaCl₂. Upper sol, 10 per cent lecithin and 5 per cent cholesterol, dispersed ultrasonically, with 0.01N NaCl and 0.001N CaCl₂.
 B. As A, but without the sodium chloride and with choline chloride 0.02N in lower sol and 0.01N in the upper sol.
 C. As A with addition of 0.01N choline chloride to both sols.
 D. As A with 0.0001N carbachol in both sols.
 E. As A with 0.00001N carbachol in both sols.
 F. As A but no lecithin or cholesterol in the upper liquid.

Under similar conditions, choline chloride has been found to diffuse rapidly. An interesting observation is that a low concentration of choline chloride (0.01N) on either side of the interface opens up the barrier to the diffusion of sodium chloride (Table I). Carbamylcholine chloride (carbachol) has a similar effect at a concentration of 0.0001N. These effects may well be related to the physiological action of choline salts in increasing the permeability of cell membranes to salts. The lipo-protein interface should provide a useful model for studying permeability changes in animal cell membranes.

L. SAUNDERS

School of Pharmacy,
 University of London,
 29/39, Brunswick Square,
 London, W.C.1.
 April 9, 1963

REFERENCE

Saunders, L. (1963). *J. Pharm. Pharmacol.*, **15**, 155-156.

SOLD

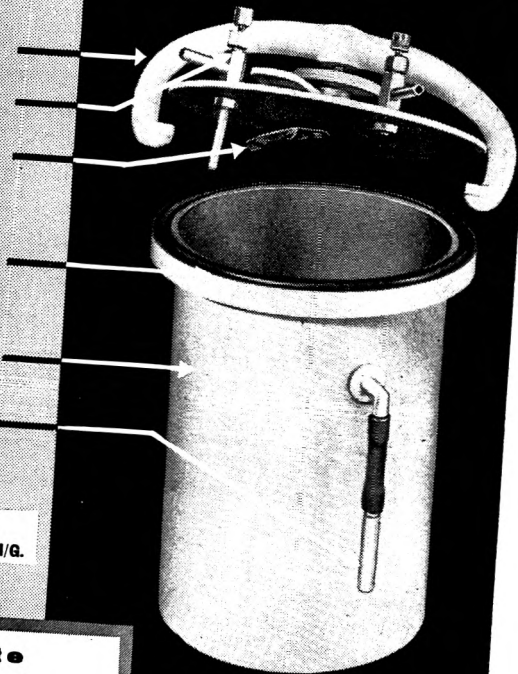
to over 4,000 satisfied buyers, this most desirable residence for anaerobes. All mod cons.— including room temperature catalyst, all metal construction, unique "o" ring sealing.

by BAIRD & TATLOCK (LONDON) LTD., CHADWELL HEATH, ESSEX, ENGLAND

Branches in London, Manchester & Glasgow.

Agents throughout the U.K. and all over the world.

- ★ Lid secured with light strong clamp.
- ★ Needle valves for connection to vacuum and hydrogen supply.
- ★ Special room temperature catalyst completely eliminates need for electrical or auxiliary heating.
- ★ 'O' ring eliminates use of grease and plasticine and gives perfect seal with hand tightness.
- ★ Strong metal jar, nickel-plated inside to resist corrosion.
- ★ External indicator contained in capsule attached to side arm.



B.T.L. Anaerobic Jar, Cat. No. D2/0050.
Price £12-12-0. Please write for leaflet SL1/G.



complete
laboratory
service

To be published in JULY

THE BRITISH PHARMACEUTICAL CODEX 1963

An international reference work providing up-to-date information on nearly 1000 medicinal substances in current use—drugs and formulated dosage forms, antisera, vaccines, etc., human-blood preparations, ligatures, sutures and surgical dressings.

Discusses the actions, uses and dosages of medicinal substances and also provides standards for those that are not included in the British Pharmacopœia.

The 1963 Codex will include about 150 entirely new monographs and 69 new formulae.

Nearly 1500 pages. Price in U.K. 105s. (plus postage 2s. 9d.)

THE PHARMACEUTICAL PRESS

17 Bloomsbury Square, London, W.C.1.

DEPARTMENT OF TECHNICAL CO-OPERATION GOVERNMENT OF TURKEY

Expert in pharmacology, and expert in analytical pharmacy, required to advise on the organisation of the Drug Control Section of the Central Institute of Hygiene in Ankara, to guide laboratory work and to give specialised training to selected staff. The present staff of the Institute is understood to consist of 6 chemists and subordinate staff.

Appointment on agreement for 2 years under British Government. Salary £3640 a year for married man; £3075 single. First £2500 in each case subject to British income tax, remainder tax free. Child and education allowance. Rent allowance. Free passages. Leave at the rate of six weeks a year.

Application forms from:

**Director of Recruitment, Department of Technical Co-operation
3 Sanctuary Buildings, Great Smith Street, London, S.W.1**

(Please quote RC205/152/06 and state full name).

EPI

Edinburgh Pharmaceutical Industries Ltd

require a **GRADUATE** to undertake work in the fields of toxicology and pharmacology. Applicants should be familiar with pharmacological techniques: previous research experience is desirable but not essential. The post offers an opportunity for a young graduate to join a research team engaged in the search for new drugs. Attractive working conditions are offered. Salary in accordance with qualifications and experience.

Applications should be sent to:

The Research Director,
Edinburgh Pharmaceutical Industries Limited,

Wheatfield Road,



Edinburgh 11.

MAKERE UNIVERSITY COLLEGE

EAST AFRICA

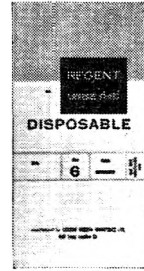
Chair in Pharmacology

Applications are invited for the newly established Chair and Headship of Department of Pharmacology.

Salaries: If medically qualified £2900 per annum; if not £2600 per annum. F.S.S.U.; Rent £45-£84 per annum according to quarters provided. Expatriation allowance £400 per annum. Education allowances for children educated outside Uganda. Up to five adult overseas passages on appointment, termination and leave (3 months every 21 months).

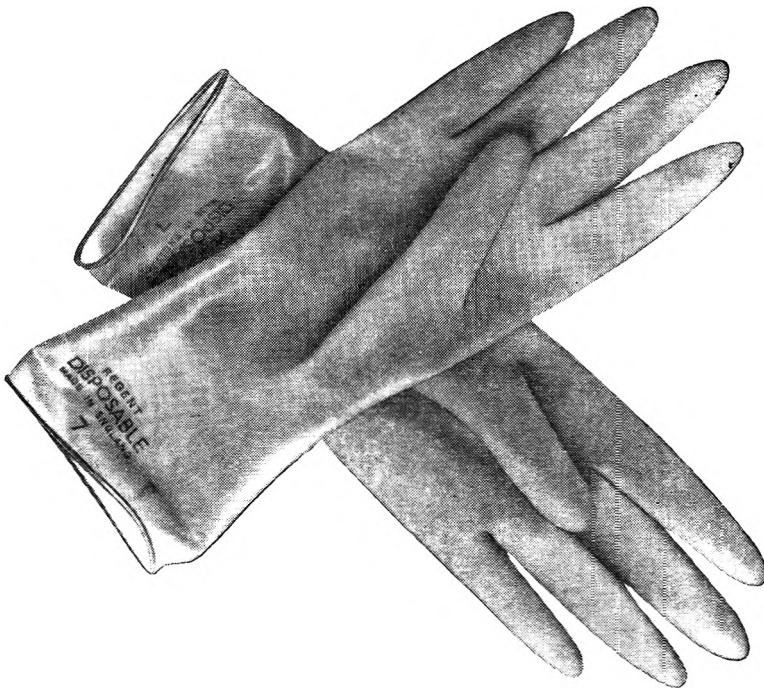
Detailed applications (8 copies) naming three referees by May 20, 1963 to Secretary, Inter-University Council for Higher Education Overseas, 29 Woburn Square, London, W.C.1., from whom further particulars may be obtained.

Save time and money with Regent Disposable Gloves by eliminating multi-stage reconditioning processes, thus simplifying sterilisation and liberating valuable skill for nursing. Because hands are assumed to be an important vehicle of cross-infection, the use of a new glove of guaranteed integrity each time also makes a contribution towards the elimination of this problem. Regent Disposable gloves reduce hand fatigue and provide 'bare hand' tactile sensitivity. They are supplied packed ready for instant sterilisation, complete with Ethicon Bio-sorb powder.



Regent Surgeons' Disposable Gloves

British Standard 1803 : 1952



Available under Ministry of Health Central Contract

Manufactured by LONDON RUBBER INDUSTRIES LIMITED
HALL LANE · LONDON E4
(Surgical Division)